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Instruction For Use 2013-10





ORG 618 Anti-PR3 hs

NAME AND INTENDED USE

Anti-PR3 hs is an ELISA test system for the quantitative measurement of IqG class autoantibodies against proteinase 3 (PR3) in human serum or plasma. This product is intended for professional in vitro diagnostic use only.

OVMBOLO HOED ON LABELO

SYMBO	DLS USED ON LABELS		
IVD	In vitro diagnostic medical device	MICROPLATE	Microplate
ш	Manufacturer	CALIBRATOR A	Calibrator
	Manufacturer	CALIBRATOR B	Calibrator
REF	Catalogue number	CALIBRATOR C	Calibrator
\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\	0.55	CALIBRATOR D	Calibrator
∑ 96	Sufficient for 96 determinations	CALIBRATOR E	Calibrator
LOT	Batch code	CALIBRATOR F	Calibrator
		CONTROL +	Control positive
$\geq \leq$	Use by	CONTROL -	Control negative
2°C 18°C	Temperature limitation		
[] i]	Consult instructions for use	DILUENT	Sample Buffer P
	Consult instructions for use	CONJUGATE	Enzyme Conjugate
类	Keep away from sunlight	ТМВ	TMD Code streets
(2)	Do not reuse	<u> </u>	TMB Substrate
	Do not reuse	STOP	Stop solution
M	Date of manufacture	WASH	Wash Buffer
		RTU	Ready to use

PRINCIPLE OF THE TEST

Highly purified proteinase 3 (PR3) is bound to microwells.

Antibodies against the coated antigen, if present in diluted patient sample, bind to the respective antigen. Washing of the microwells removes unbound unspecific serum and plasma components. Horseradish peroxidase (HRP) conjugated anti-human antibodies immunologically detect the bound patient antibodies forming a conjugate/antibody/antigen complex. Washing of the microwells removes unbound conjugate. An enzyme substrate in the presence of bound conjugate hydrolyzes to form a blue colour. The addition of an acid stops the reaction forming a yellow end-product. The intensity of this yellow colour is measured photometrically at 450 nm. The amount of colour is directly proportional to the concentration of antibodies present in the original sample.

SUMMARY AND EXPLANATION OF THE TEST

Anti-neutrophil cytoplasmic antibodies (ANCA) represent a group of autoantibodies directed towards cytoplasmic components of the neutrophil granulocytes and monocytes. The classical methods for the determination of ANCA are immunofluorescence tests. With these indirect immunofluorescence (IF) techniques two main patterns are distinguished: a cytoplasmic (cANCA) and a perinuclear (pANCA) type.

The target antigen for 80-90 % of cANCA is proteinase 3 (PR3), a serine proteinase present in primary granules: 10 -20 % of cANCA are directed to other proteins, such as bactericidal permeability-increasing protein (BPI). In rare cases, antibodies to elastase (4 %), lysozym (2 %) or cathepsin G (2 %) may show a cANCA-pattern. cANCA have also been detected in different non-rheumatic diseases.

Approximately 90 % of pANCA positive sera contain autoantibodies directed to myeloperoxidase (MPO), which is located in the granules of neutrophil granulocytes. Antibodies to other antigens e.g. Lactoferrin, Elastase. Cathepsin-G and also Lysozyme often result in a similar pANCA pattern. These atvoical pANCA occur in collagenosis and related inflammatory rheumatic diseases. Besides, different untypical variants of pANCA IF patterns - granulocyte specific antinuclear antibodies (GS-ANA) - are indistinguishable from pANCA.

Therefore, a distinct interpretation and classification of the IF patterns is difficult and every positive IF-ANCA finding should be differentiated by ELISA techniques using the purified single antigens.

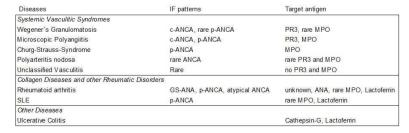
PR3 and MPO are well defined and reliable serologic markers for a definite group of primary systemic vasculitides (PSV), they are also called ANCA-associated vasculitides (AAV). The incidence is 1 in 1000 in the whole population and nearly 5 in 1000 in the elderly. The clinical appearance of AAV is characterised by manifestations in the kidneys, the lung and the respiratory tract.

PR3 is the most frequent component of cANCA and the landmark autoantiqen in granulomatosis with polyangiitis (GPA, formerly named Wegener's granulomatosis) with a clinical specificity of more than 95% for the disease.

MPO, the main target antigen of pANCA, is present in 70% of patients with microscopic polyangiitis (MPA) and differentiates MPA from other autoimmune diseases.

Anti-PR3 and anti-MPO levels correlate with the clinical status; they are high in active disease. Antibody titres decrease under therapy and become undetectable after remission.

A survey of documented clinical indications, the corresponding immunofluorescence patterns and target antigens is given in the following table:



CONTENTS OF THE KIT

ORG 618	₩ 96	Sufficient for 96 determinations
MICROPLATE	1	One divisible microplate consisting of 12 modules of 8 wells each. Ready to use. Product code on module: PR3
CALIBRATOR A	1x 1.5 ml	Calibrator A 0 U/ml, containing serum/buffer matrix (PBS, BSA, detergent, NaN3 0.09%), yellow. Ready to use.
CALIBRATOR B	1x 1.5 ml	Calibrator B 10 U/ml, containing PR3 antibodies in a serum/buffer matrix (PBS, BSA, detergent, NaN3 0.09%), yellow. Ready to use.
CALIBRATOR C	1x 1.5 ml	Calibrator C 20 U/ml, containing PR3 antibodies in a serum/buffer matrix (PBS, BSA, detergent, NaN3 0.09%), yellow. Ready to use.
CALIBRATOR D	1x 1.5 ml	Calibrator D 40 U/ml, containing PR3 antibodies in a serum/buffer matrix (PBS, BSA, detergent, NaN3 0.09%), yellow. Ready to use.
CALIBRATOR E	1x 1.5 ml	Calibrator E 80 U/ml, containing PR3 antibodies in a serum/buffer matrix (PBS, BSA, NaN3 0.09%), yellow. Ready to use.
CALIBRATOR F	1x 1.5 ml	Calibrator F 200 U/ml, containing PR3 antibodies in a serum/buffer matrix (PBS, BSA, detergent, NaN3 0.09%), yellow. Ready to use.
CONTROL +	1x 1.5 ml	Control positive, containing PR3 antibodies in a serum/buffer matrix (PBS, BSA, detergent, NaN3 0.09%), yellow. Ready to use. The concentration is specified on the certificate of analysis.
CONTROL -	1x 1.5 ml	Control negative, containing PR3 antibodies in a serum/buffer matrix (PBS, BSA, detergent, NaN3 0.09%), yellow. Ready to use. The concentration is specified on the certificate of analysis.
DILUENT	20 ml	Sample Buffer P, containing PBS, BSA, detergent, preservative sodium azide 0.09% , yellow, concentrate (5 x).
CONJUGATE	15 ml	Enzyme Conjugate containing anti-human IgG antibodies, HRP labelled; PBS, BSA, detergent, preservative PROCLIN 0.05%, light red. Ready to use.
ТМВ	15 ml	TMB Substrate; containing 3,3', 5,5'- Tetramethylbenzidin, colorless. Ready to use.
STOP	15 ml	Stop solution; contains acid. Ready to use.
WASH	20 ml	Wash Buffer, containing Tris, detergent, preservative sodium azide 0.09%; 50 x conc.
[]i	1	Instruction for Use: ELISA Mini-DVD
Ti	1	Certificate of Analysis

MATERIALS REQUIRED

- · Microplate reader capable of endpoint measurements at 450 nm; optional: reference filter at 620 nm
- · Data reduction software
- Multi-channel dispenser or repeatable pipette for 100 µl
- Vortex mixer
- Pipettes for 10 μl, 100 μl and 1000 μl
- · Laboratory timing device
- · Distilled or deionised water
- · Measuring cylinder for 1000 ml and 100 ml
- · Plastic container for storage of the wash solution

This ELISA assay is suitable for use on open automated ELISA processors. Each assay has to be validated on the respective automated system. Detailed information is provided upon request.

SPECIMEN COLLECTION, STORAGE AND HANDLING

- · Collect whole blood specimens using acceptable medical techniques to avoid hemolysis.
- Allow blood to clot and separate the serum or plasma by centrifugation.
- Test serum should be clear and non-hemolyzed. Contamination by hemolysis or lipemia should be avoided, but does not interfere with this assay.
- Specimens may be refrigerated at 2-8°C for up to five days or stored at -20°C up to six months.
- Avoid repetitive freezing and thawing of serum or plasma samples. This may result in variable loss of antibody activity.
- · Testing of heat-inactivated sera is not recommended.

STORAGE AND STABILITY

- . Store test kit at 2-8°C in the dark.
- Do not expose reagents to heat, sun, or strong light during storage and usage.
- · Store microplate sealed and dessicated in the clip bag provided.
- Shelf life of the unopended test kit is 18 months from day of production.
 Unopened reagents are stable until expiration of the kit. See labels for individual batch.
- Diluted Wash Buffer and Sample Buffer are stable for at least 30 days when stored at 2-8°C.
 We recommend consumption on the same day.

PROCEDURAL NOTES

- Do not use kit components beyond their expiration dates.
- Do not interchange kit components from different lots and products.
- All materials must be at room temperature (20-28°C) prior to use.
- · Prepare all reagents and samples. Once started, performe the test without interruption.
- Double determinations may be done. By this means pipetting errors may become obvious.
- · Perform the assay steps only in the order indicated.
- · Always use fresh sample dilutions.
- · Pipette all reagents and samples into the bottom of the wells.
- To avoid carryover or contamination, change the pipette tip between samples and different kit controls.
- · Wash microwells thoroughly and remove the last droplets of wash buffer.
- · All incubation steps must be accurately timed.
- · Do not re-use microplate wells.

WARNINGS AND PRECAUTIONS

- · All reagents of this kit are intended for professional in vitro diagnostic use only.
- Components containing human serum were tested and found negative for HBsAg, HCV, HIV1 and HIV2 by FDA approved methods. No test can guarantee the absence of HBsAg, HCV, HIV1 or HIV2, and so all human serum based reagents in this kit must be handled as though capable of transmitting infection.
- Bovine serum albumin (BSA) used in components has been tested for BSE and found negative.
- Avoid contact with the substrate TMB (3,3',5,5'-Tetramethyl-benzidine).
- Stop solution contains acid, classifiaction is non-hazardous. Avoid contact with skin.
- Control, sample buffer and wash buffer contain sodium azide 0.09% as preservative. This concentration is classified as non-hazardous.
- Enzyme conjugate contains ProClin 300 0.05% as preservative. This concentration is classified as non-hazardous.

During handling of all reagents, controls and serum samples observe the existing regulations for laboratory safety regulations and good laboratory practice:

- First aid measures: In case of skin contact, immediately wash thoroughly with water and soap. Remove
 contaminated clothing and shoes and wash before reuse. If system fluid comes into contact with skin,
 wash thoroughly with water. After contact with the eyes carefully rinse the opened eye with running
 water for at least 10 minutes. Get medical attention if necessary.
- Personal precautions, protective equipment and emergency procedures:

Observe laboratory safety regulations. Avoid contact with skin and eyes. Do not swallow. Do not pipette by mouth. Do not eat, drink, smoke or apply makeup in areas where specimens or kit reagents are handled. When spilled, absorb with an inert material and put the spilled material in an appropriate waste disposal.

- Exposure controls / personal protection: Wear protective gloves of nitril rubber or natural latex. Wear protective glasses. Used according to intended use no dangerous reactions known.
- · Conditions to avoid: Since substrate solution is light-sensitive. Store in the dark.
- · For disposal of laboratory waste the national or regional legislation has to be observed.

Observe the guidelines for performing quality control in medical laboratories by assaying control sera.

PREPARATION OF REAGENTS

WASH

Dilute the contents of one vial of the buffered wash solution concentrate (50x) with distilled or deionised water to a final volume of 1000 ml prior to use.

DILUENT

Sample Buffer P: Prior to use dilute the contents (20 ml) of one vial of sample buffer 5x concentrate with distilled or deionised water to a final volume of 100 ml.

Preparation of samples

Dilute all patient samples **1:200** with sample buffer prior to use in the assay: combine **5 µI** of sample with 995 µI of sample buffer in a polystyrene tube. Mix well. Note: Calibrators / Controls need no dilution.

TEST PROCEDURE

Prepare enough microplate modules for all calibrators / controls and patient samples.

1. Pipette 100 μI of calibrators, controls and prediluted patient samples into the wells.

Incubate for 30 minutes at room temperature (20-28 °C).

Discard the contents of the microwells and wash 3 times with 300 µl of wash solution.

2. Dispense 100 ul of enzyme conjugate into each well.

Incubate for 15 minutes at room temperature.

Discard the contents of the microwells and wash 3 times with 300 µl of wash solution.

3. Dispense 100 ul of TMB substrate solution into each well

Incubate for 15 minutes at room temperature

4. Add 100 µl of stop solution to each well of the modules

Incubate for 5 minutes at room temperature.

Read the optical density at 450 nm (reference 600-690nm) and calculate the results.

The developed colour is stable for at least 30 minutes. Read during this time.

Example for a pipetting scheme:

	1	2	3	4	5	6	7	8	9	10	11	12
Α	Α	P1										
В	В	P2										
С	C	P3										
D	D											
Е	Е											
F	F											
G	C+											
Н	C-											

P1, ... patient sample A-F calibrators C+, C- controls

VALIDATION

Test results are valid if the optical densities at 450 nm for calibrators / controls and the results for controls comply with the reference ranges indicated on the Certificate of Analysis enclosed in each test kit.

If these quality control criteria are not met the assay run is invalid and should be repeated.

CALCULATION OF RESULTS

For quantitative results plot the optical density of each calibrator versus the calibrator concentration to create a calibration curve. The concentration of patient samples may then be estimated from the calibration curve by interpolation.

Using data reduction software a 4-Parameter-Fit with lin-log coordinates for optical density and concentration is the data reduction method of choice.

PERFORMANCE CHARACTERISTICS

CALIBRATION

This assay system is calibrated in relative arbitrary units, since no international reference preparation is available for this assay.

Measuring range

The calculation range of this ELISA assay is 0 - 200 U/ml

Expected values

In a normal range study with samples from healthy blood donors the following ranges have been established with this ELISA assav: Cut-off 10 U/ml

Interpretation of results

Negative: < 10 U/ml Positive: ≥ 10 U/ml

Linearity

Patient samples containing high levels of specific antibody were serially diluted in sample buffer to demonstrate the dynamic range of the assay and the upper / lower end of linearity. Activity for each dilution was calculated from the calibration curve using a 4-Parameter-Fit with lin-log coordinates.

Sample	Dilution	Observed	Expected	O/E
		U/ml	U/ml	[%]
1	1:100	<mark>168.2</mark>	168.2	100
	1:200	84.6	84.1	101
	1:400	41.3	42.1	98
	1:800	20.1	21.0	96
	1:1600	9.7	10.5	92
2	1:100	122.1	122.1	100
	1:200	60.7	61.1	99
	1:400	29.8	30.5	98
	1:800	14.6	15.3	95
	1:1600	<mark>6.7</mark>	7.6	88

Limit of detection

Functional sensitivity was determined to be: 0.5 U/ml

Reproducibility

Intra-assay precision: Coefficient of variation (CV) was calculated for each of three samples from the results of 24 determinations in a single run. Results for precision-within-assay are shown in the table below.

Inter-assay precision: Coefficient of variation (CV) was calculated for each of three samples from the results of 6 determinations in 5 different runs. Results for run-to-run precision are shown in the table below.

Intra-Assay					
Sample					
	U/ml	CV %			
1	11.9	3.5			
2	44.7	2.0			
3	132.8	2.5			

Inter-Assay						
Sample	Mean					
	U/ml	CV %				
1	12.6	3.8				
2	46.8	4.1				
3	122.7	4.2				

Interfering substances

No interference has been observed with haemolytic (up to 1000 mg/dl) or lipemic (up to 3 g/dl triglycerides) sera or plasma, or bilirubin (up to 40 mg/dl) containing sera or plasma. Nor have any interfering effects been observed with the use of anticoagulants (Citrate, EDTA, Heparine). However for practical reasons it is recommended that grossly hemolyzed or lipemic samples should be avoided.

Study results

Study population	<u>n</u>	n Pos	%
Morbus Wegener (cANCA pos)	51	49	96.1
inflam./Non-inflammatory disorders	450	8	1.8
Healthy controls	80	0	0.0

Immunological Diagnosis

		POS	iveg	
ORG 618	Pos	49	8	
	Neg	2	522	
		51	530	581

Sensitivity: 96.1 % Specificity: 98.5 % Overall agreement: 98.3 %

LIMITATIONS OF THE PROCEDURE

This assay is a diagnostic aid. A definite clinical diagnosis should not be based on the results of a single test, but should be made by the physician after all clinical and laboratory findings have been evaluated concerning the entire clinical picture of the patient. Also every decision for therapy should be taken individually.

The above pathological and normal reference ranges for antibodies in patient samples should be regarded as recommendations only. Each laboratory should establishe its own ranges according to ISO 15189 or other applicable laboratory guidelines.

REFERENCES

- Cohen Tervaert J.W., Goldschmeding, R., Elema, J.D., Limburg, P.C. et al Association of autoantibodies to myeloperoxidase with different forms of vasculitis. Arthritis Rheum. 33 (8):1264-1272, 1990
- Cotch, M.F., Hoffman, G.S., Yerg, D.E., et.al. The epidemiology of Wegener's granulomatosis. Estimates of the five-year period prevalence, annual mortality, and geographic disease distribution from population-based data sources. Arthritis Rheum., Vol 39, 87-92, 1996
- de Grot, K., Schnabel, A., Gross, W. L. ANCA-assoziierte Vaskulitiden (Wegener Granulomatose, Churg-Strauß Syndrom, Mikroskopische Polyanqiitis) - 1. Diagnostisches Procedere. Z. Rheumatol., Vol 54, 291 - 302, 1995
- 4. Gross, W.L. Systemic necrotizing vasculitis. Baillieres. Clin. Rheumatol., Vol. 11, 259-284, 1997
- Gross, W.L., Csernok, E., Helmchen, U. Antineutrophil cytoplasmic autoantibodies, autoantigens, and systemic vasculitis. APMIS, Vol. 103, 81-97, 1995
- Jenette, J. C. Falk, R. J. Antineutrophil cytoplasmic autoantibodies and associated diseases. A review. Am. J. Kidney Dis., Vol 15, 517 - 529, 1990
- 7. Jennette, C.J., Falk, R.J., Andrassy, K., Bacon, P.A. et al. Nomenclature of systemic vasculitides. Proposal of an international consensus conference. Arthritis Rheum. Vol. 37, 187-192, 1994
- 8. Kallenberg, C.G Vasculitis 1995: new concepts in pathophysiology. Isr.J Med Sci., Vol. 32, 7-12, 1996
- 9. Kallenberg, C.G. Laboratory findings in the vasculitides. Baillieres Clin Rheumatol., Vol 11, 395-421, 1997
- Schmitt, W. H., Gross, W. L. Antineutrophile zytoplasmatische Antikörper (ANCA). Internist, Vol. 36, 282 290, 1995
- 11.Wiik, A., Stumman, L., Kjeldsen, L., Borregaard, N., et al. The diversity of perinuclear antineutrophil cytoplasmic antibodies (pANCA) antigens. Clin. Exp. Immunol., Vol. 101, Suppl. I, 15-17, 1995

Pipet 100 µl calibrator, control or patient sample
Incubate for 30 minutes at room temperature

Discard the contents of the wells and wash 3 times with 300 µl wash solution

Pipet 100 µl enzyme conjugate

Incubate for 15 minutes at room temperature

Discard the contents of the wells and wash 3 times with 300 µl wash solution

Pipet 100 µl substrate solution

Incubate for 15 minutes at room temperature

Add 100 µl stop solution

Leave untouched for 5 minutes

Read at 450 nm