

This kit is intended for Research Use Only.

Not for use in diagnostic procedures.

Store at 2 to 8°C.

1. SAMPLE DILUTION 1:40

5 µl / 200 µl

2. Three incubations at 37°C

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| Diluted Sample 100 µl 30 min. | Enzyme Conjugate 100 µl 30 min. | TMB Reagent (One-Step) 100 µl 15 min. |
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3. STOP WITH 100 µL OF ACID. READ O.D. AT 450 NM

INTENDED USE

The HSV-2 IgG ELISA is intended for use in measuring serologic status to herpes simplex virus (HSV) type 2 infection, or for evaluating paired sera for the presence of a significant increase in herpes specific IgG.

PRINCIPLE OF THE TEST

Purified HSV-2 antigen is coated on the surface of microwells. Diluted serum is added to wells, and the HSV-2 IgG-specific antibody, if present, binds to the antigen. All unbound materials are washed away. HRP-conjugate is added, which binds to the antibody-antigen complex. Excess enzyme conjugate is washed off and a solution of TMB Reagent is added. The enzyme conjugate catalytic reaction is stopped at a specific time. The intensity of the color generated is proportional to the amount of HSV-2 IgG-specific antibody in the sample. The results are read by a microwell reader compared in a parallel manner with calibrator and controls.

REAGENTS

Materials provided with the kit:

- Microtiter Wells: purified HSV-2 antigen coated wells (12 x 8 wells)
- Enzyme Conjugate Reagent (red color): 1 vial (12 ml)
- Sample Diluent (green color): 1 bottle (22 ml)
- Negative Control: Range stated on label. Natural cap (100 µL/vial)
- Cut-off Calibrator: Yellow cap. HSV-2 IgG Index = 1 (100 µL/vial)
- Positive Control: Range stated on label. Red cap. (100 µL/vial)
- Wash Buffer Concentrate (20x): 1 bottle (50 ml)
- TMB Reagent (One-Step): 1 vial (11 ml)
- Stop Solution (1N HCl): Natural cap. 1 vial (11 ml)

STORAGE OF TEST KITS AND INSTRUMENTATION

1. Store the kit at 2-8°C.
2. Keep microwells sealed in a dry bag with desiccants.

3. The reagents are stable until expiration of the kit.
4. Do not expose test reagents to heat, sun or strong light during storage or usage.

WARNING AND PRECAUTIONS

1. Potential biohazardous materials:
The calibrator and controls contain human source components which have been tested and found non-reactive for hepatitis B surface antigen as well as HIV antibody with FDA licensed reagents. However, as there is no test method that can offer complete assurance that HIV, hepatitis B virus or other infectious agents are absent, these reagents should be handled at the Biosafety Level 2, as recommended in the Centers for Disease Control/National Institutes of Health manual, "Biosafety in Microbiological and Biomedical Laboratories." 1984
2. Do not pipette by mouth. Do not smoke, eat, or drink in the areas in which specimens or kit reagents are handled.
3. The components from different lots should not be mixed.
4. This product contains components preserved with sodium azide. Sodium azide may react with lead and copper plumbing to form explosive metal azide. On disposal, flush with a large volume of water.

SPECIMEN COLLECTION AND PREPARATION

1. Collect blood specimens and separate the serum.
2. Specimens may be refrigerated at 2-8°C for up to 7 days or frozen for up to 6 months. Avoid repetitive freezing and thawing of serum sample.

REAGENT PREPARATION

1. All reagents should be allowed to reach room temperature (18-25 °C) before use.
2. Dilute 1 volume of Wash Buffer (20x) with 19 volumes of distilled water. For example, dilute 50 ml of Wash Buffer (20x) into distilled water to prepare 1000 ml of Wash Buffer (1x). Wash Buffer is stable for 1 month at 2-8°C. Mix well before use.

ASSAY PROCEDURE

1. Place the desired number of coated wells into the holder.
2. Prepare 1:40 dilution of test samples, Negative Control, Positive Control, and Calibrator by adding 5 µl of the sample to 200 µl of Sample Diluent. Mix well.
3. Dispense 100 µl of diluted sera, Calibrator, and Controls into the appropriate wells. For the reagent blank, dispense 100 µl Sample Diluent in 1A well position. Tap the holder to remove air bubbles from the liquid and mix well.
4. Incubate at 37°C for 30 minutes.
5. At the end of incubation period, remove liquid from all wells. Rinse and flick the microtiter wells 5 times with diluted Wash Buffer (1x).
6. Dispense 100 µl of Enzyme Conjugate to each well. Mix gently for 10 seconds.
7. Incubate at 37°C for 30 minutes.
8. Remove Enzyme Conjugate from all wells. Rinse and flick the microtiter wells 5 times with diluted Wash Buffer (1x).
9. Dispense 100 µl of TMB Reagent into each well. Mix gently for 10 seconds.

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10. Incubate at 37°C for 15 minutes.
11. Add 100 µl of Stop Solution to stop reaction.
12. Mix gently for 30 seconds. *It is important to make sure that all the blue color changes to yellow color completely.*

Note: Make sure there are no air bubbles in each well before reading.
Read O.D. at 450nm *within 15 minutes* with a microwell reader.

CALCULATION OF RESULTS

1. Calculate the mean of duplicate calibrator value x_c .
2. Calculate the mean of duplicate positive control (x_p), negative control (x_n) and samples (x_s).
3. Calculate the HSV-2 IgG Index of each determination by dividing the mean values of each sample (x) by calibrator mean value, x_c .

Example of typical results: Note: The O.D. values are for the purpose of illustration only, and should not be used to calculate unknowns. Each user should obtain his or her own data.

Cut-off Calibrator HSV-2 IgG Index = 1.0

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| Cut-off Calibrator O.D. = 0.889, 0.864 | $x_c = 0.877$ |
| 2. Negative Control O.D. = 0.077, 0.083 | $x_n = 0.080$ |
| HSV-2 IgG Index = $x_n / x_c = 0.080 / 0.877 = 0.09$ | |
| 3. Positive Control O.D. = 1.645, 1.668 | $x_p = 1.657$ |
| HSV-2 IgG Index = $x_p / x_c = 1.657 / 0.877 = 1.89$ | |
| 4. Donor sample O.D. = 3.087, 3.144 | $x_s = 3.116$ |
| HSV-2 IgG Index = $x_s / x_c = 3.116 / 0.877 = 3.55$ | |

QUALITY CONTROL

The test run may be considered valid provided the following criteria are met:

1. The O.D. value of the reagent blank against air from a microwell reader should be less than 0.250.
2. If the O.D. value of the Cut-off Calibrator is lower than 0.250, the test is not valid and must be repeated.
3. The HSV-2 IgG Index for Negative and Positive Control should be in the range stated on the Certificate of Analysis.

LIMITATIONS OF THE PROCEDURE

1. Reliable results will be obtained when the assay procedure is carried out with a complete understanding of the package insert instructions and with adherence to good laboratory practice.
2. The wash procedure is critical. Insufficient washing will result in poor precision and falsely elevated absorbance readings.
3. Serum samples demonstrating gross lipemia, gross hemolysis, or turbidity should not be used with this test.

REFERENCES

1. Nahmias, A.J., J. Dannenbarger, C. Wickliffe and M. Muther. Clinical aspects of infection with herpes simplex viruses 1 and 2 in the human herpes viruses. An interdisciplinary Perspective (Nahmias, A.J., W.R. Dawdle and R.F. Schinazi eds) New York, Elsevier, pp 3-9, 1981.
2. Vestergaard, B.F., P.C. Grauballe and H. Spanggaard. Titration of herpes simplex virus antibodies in human sera by the enzyme-link immunosorbent assay (ELISA). Acta Pathol. Microbiol. Scand. Sect. B 85:446-448, 1977.
3. Coleman, R.M., L. Pereira, P.D. Bailey, D. Dondero, C. Wickliffe, and A.J. Nahmias. Determination of herpes simplex virus type-specific antibodies by enzyme-linked immunosorbent assay. J. Clin. Microbiol. 18 (1983) 287.

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