AESKULISA Crohn's Check

REF 3509

Instruction manual

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002 : 2007-08-28

1. Intended Use

AESKULISA Crohn`s-Check is a solid phase enzyme immunoassay (ELISA) employing highly purified mannan for the combined quantitative and qualitative detection of anti-Saccharomyces cerevisiae IgA and IgG antibodies (ASCA) in human serum.

ASCA recognize specific mannan, a component of the outer cell wall of yeast. The assay is highly specific and sensitive for Crohn's disease.

2. Clinical Application and Principle of the Assay

Crohn's disease is one of the two major Inflammatory Bowel Diseases (IBD). IBD is an umbrella term, that covers both primary disorders causing inflammation or ulceration in the small and large intestine, Crohn's disease and ulcerative colitis. Crohn's disease affects both, the small bowel and the colon, unlike ulcerative colitis which is restricted to the colon only. The etiology is not revealed yet, although a genetic and infectious background for the disease is under discussion. Colonoscopy and ileoscopy are the established tools of diagnosis, no serology was available so far. Though Crohn's disease and ulcerative colitis share several symptoms the course of the diseases, its complications and its management are different, especially when it comes down to surgery. Consequently the differental diagnosis of both diseases is crucial prior to treatment. Aggravating, about 5-10% of the patients can not be distinguished clearly by existing available diagnostic methodologies and are referred to as indeterminate colitis.

ASCA have been found to be specific markers for Crohn's disease. They have been reported for these patients with a frequency of 68%. The identification of the target antigen mannan, a mannose-rich carbohydrate antigen of the outer cell wall of yeast, enabled the detection of ASCA by enzyme immunoassay.

Being the first available highly specific serological marker, ASCA may become an important tool for the difficult task of diagnosing IBD. Moreover its high positive predictive value offers the only possibility of a convenient and reliable screening and monitoring of risk groups.

Principle of the test

Serum samples diluted 1:101 are incubated in the microplates coated with the specific antigen. Patient's antibodies, if present in the specimen, bind to the antigen. The unbound fraction is washed off in the following step. Afterwards anti-human immunoglobulins conjugated to horseradish peroxidase (conjugate) are incubated and react with the antigen-antibody complex of the samples in the microplates. Unbound conjugate is washed off in the following step. Addition of TMB-substrate generates an enzymatic colorimetric (blue) reaction, which is stopped by diluted acid (color changes to yellow). The rate of color formation from the chromogen is a function of the amount of conjugate bound to the antigen-antibody complex and this is proportional to the initial concentration of the respective antibodies in the patient sample.

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3. Kit Contents

To be reconstituted:

5x Sample Buffer 1 vial, 20 ml - 5x concentrated (capped white: yellow solution)

Containing: Tris, NaCl, BSA, sodium azide < 0.1% (preservative)

50x Wash Buffer 1 vial, 20 ml - 50x concentrated (capped white: green solution)

Containing: Tris, NaCl, Tween 20, sodium azide < 0.1% (preservative)

Ready to use:

Negative Control 1 vial, 1.5 ml (capped green:colorless solution)

Containing: Human serum (diluted), sodium azide < 0.1% (preservative)

Positive Control 1 vial, 1.5 ml (capped red: yellow solution)

Containing: Human serum (diluted), sodium azide < 0.1% (preservative)

Cut-off Calibrator 1 vial, 1.5 ml (capped blue: yellow solution)

Containing: Human serum (diluted), Sodium azide < 0.1% (preservative)

Calibrators 6 vials, 1.5 ml each 0, 3, 10, 30, 100, 300 U/ml

(color increasing with concentration: yellow solutions)

Containing: Human serum (diluted), sodium azide < 0.1% (preservative)

Conjugate 1 vial,15 ml IgA/G (capped white: red solution)

Containing: Anti-human immunoglobulins conjugated to horseradish peroxidase

TMB Substrate 1 vial, 15 ml (capped black)

Containing: Stabilized TMB/H₂O₂

Stop Solution 1 vial, 15 ml (capped white: colorless solution)

Containing: 1M Hydrochloric Acid

Microtiterplate 12x8 well strips with breakaway microwells

Coating see paragraph 1

Material required but not provided:

Microtiter plate reader 450 nm reading filter and optional 620 nm reference filter (600-690 nm). Glass ware(cylinder 100-1000ml), test tubes for dilutions. Vortex mixer, precision pipettes (10, 100, 200, 500, 1000 μ l) or adjustable multipipette (100-1000ml). Microplate washing device (300 μ l repeating or multichannel pipette or automated system), adsorbent paper.

Our tests are designed to be used with purified water according to the definition of the United States Pharmacopeia (USP 26 - NF 21) and the European Pharmacopeia (Eur.Ph. 4th ed.).

4. Storage and Shelf Life

Store all reagents and the microplate at 2-8°C/35-46°F, in their original containers. Once prepared, reconstituted solutions are stable for 1 month at 4°C/39°F, at least. **Reagents and the microplate shall be used within the expiry date indicated on each component, only. Avoid intense exposure of TMB solution to light. Store microplates in designated foil, including the desiccant, and seal tightly.**

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5. Precautions of Use

5.1 Health hazard data

This product is for **IN VITRO DIAGNOSTIC USE** only. Thus, only staff trained and specially advised in methods of in vitro diagnostics may perform the kit. Although this product is not considered particularly toxic or dangerous in conditions of normal use, refer to the following for maximum safety:

Recommendations and precautions

This kit contains potentially hazardous components. Though kit reagents are not classified being irritant to eyes and skin we recommend to avoid contact with eyes and skin and wear disposable gloves.

WARNING! Calibrators, Controls and Buffers contain sodium azide (NaN₃) as a preservative. NaN₃ may be toxic if ingested or adsorbed by skin or eyes. NaN₃ may react with lead and copper plumbing to form highly explosive metal azides. On disposal, flush with a large volume of water to prevent azide build-up. Please refer to decontamination procedures as outlined by CDC or other local/national guidelines.

Do not smoke, eat or drink when manipulating the kit.

Do not pipette by mouth.

All human source material used for some reagents of this kit (controls, standards e.g.) has been tested by approved methods and found negative for HbsAg, Hepatitis C and HIV 1. However, no test can guarantee the absence of viral agents in such material completely. Thus handle kit controls, standards and patient samples as if capable of transmitting infectious diseases and according to national requirements.

5.2 General directions for use

Do not mix or substitute reagents or microplates from different lot numbers. This may lead to variations in the results.

Allow all components to reach room temperature (20-32°C/68-89.6°F) before use, mix well and follow the recommended incubation scheme for an optimum performance of the test.

Incubation: We recommend test performance at 30°C/86°F for automated systems.

Never expose components to higher temperature than 37°C/98.6 °F.

Always pipette substrate solution with brand new tips only. Protect this reagent from light. Never pipette conjugate with tips used with other reagents prior.

A definite clinical diagnosis should not be based on the results of the performed test only, but should be made by the physician after all clinical and laboratory findings have been evaluated. The diagnosis is to be verified using different diagnostic methods.

6. Sample Collection, Handling and Storage

Use preferentially freshly collected serum samples. Blood withdrawal must follow national requirements.

Do not use icteric, lipemic, hemolysed or bacterially contaminated samples. Sera with particles should be cleared by low speed centrifugation (<1000 x g). Blood samples should be collected in clean, dry and empty tubes. After separation, the serum samples should be used immediately, respectively stored tightly closed at $2-8^{\circ}\text{C}/35-46^{\circ}\text{F}$ up to three days, or frozen at $-20^{\circ}\text{C}/-4^{\circ}\text{F}$ for longer periods.

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7. Assay Procedure

7.1 Preparations prior to pipetting

Dilute concentrated reagents:

Dilute the concentrated sample buffer 1:5 with distilled water (e.g. 20 ml plus 80 ml). Dilute the concentrated wash buffer 1:50 with distilled water (e.g. 20 ml plus 980 ml).

Samples:

Dilute serum samples 1:101 with sample buffer (1x) e.g. 1000 µl sample buffer (1x) + 10 µl serum. Mix well!

Washing:

Prepare 20 ml of diluted wash buffer (1x) per 8 wells or 200 ml for 96 wells e.g. 4 ml concentrate plus 196 ml distilled water.

Automated washing:

Consider excess volumes required for setting up the instrument and dead volume of robot pipette.

Manual washing:

Discard liquid from wells by inverting the plate. Knock the microwell frame with wells downside vigorously on clean adsorbent paper. Pipette 300 µl of diluted wash buffer into each well, wait for 20 seconds. Repeat the whole procedure twice again.

Microplates:

Calculate the number of wells required for the test. Remove unused wells from the frame, replace and store in the provided plastic bag, together with desiccant, seal tightly (2-8°C/35-46°F).

7.2 Work flow

For pipetting scheme see Annex A, for the test procedure see Annex B We recommend pipetting samples and calibrators in duplicate.

Cut-off calibrator should be used for qualitative testing only.

- Pipette 100 μl of each patient's diluted serum into the designated microwells.
- Pipette 100 µl calibrators OR cut-off calibrator and negative and positive controls into the designated wells.
- Incubate for 30 minutes at 20-32°C/68-89.6°F.
- Wash 3x with 300 µl washing buffer (diluted 1:50).
- Pipette 100 µl conjugate into each well.
- Incubate for 30 minutes at 20-32°C/68-89.6°F.
- Wash 3x with 300 µl washing buffer (diluted 1:50).
- Pipette 100 µl TMB substrate into each well.
- Incubate for 30 minutes at 20-32°C/68-89.6°F, protected from intense light.
- Pipette 100 µl stop solution into each well, using the same order as pipetting the substrate.
- Incubate 5 minutes minimum.
- Agitate plate carefully for 5 sec.
- Read absorbance at 450 nm (optionally 450/620 nm) within 30 minutes.

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8. Quantitative and Qualitative Interpretation

For quantitative interpretation establish the standard curve by plotting the optical density (OD) of each calibrator (y-axis) with respect to the corresponding concentration values in U/ml (x-axis). For best results we recommend log/lin coordinates and 4-Parameter Fit. From the OD of each sample, read the corresponding antibody concentrations expressed in U/ml.

Normal Range	Equivocal Range	Positive Results
< 16 U/ml	16 - 24 U/ml	>24 U/ml

Example of a standard curve

We recommend pipetting calibrators in parallel for each run.

Calibrators IgA/G	OD 450/620 nm	CV % (Variation)
0 U/ml	0.032	2.8
3 U/ml	0.152	2.6
10 U/ml	0.281	1.2
30 U/ml	0.646	2.4
100 U/ml	1.214	1.7
300 U/ml	2.104	1.6

Example of calculation

Patient	nt Replicate (OD) Mean (OD)		Result (U/ml)
P 01	0.904/0.937	0.921	58.3
P 02	0.564/0.551	0.558	25.9

For lot specific data, see enclosed quality control leaflet. Medical laboratories might perform an inhouse Quality Control by using own controls and/or internal pooled sera, as foreseen by EU regulations.

Do not use this example for interpreting patients results!

Each laboratory should establish its own normal range based upon its own techniques, controls, equipment and patient population according to their own established procedures.

For qualitative interpretation read the optical density of the cut-off calibrator and the patient samples. Compare patient'sOD with the OD of the cut-off calibrator. For qualitative interpretation we recommend to consider sera within a range of 20% around the cut-off value as equivocal. All samples with higher ODs are considered positive, samples with lower ODs are considered negative.

Negative: OD patient < 0.8 x OD cut-off

Equivocal: $0.8 \times OD_{cut-off} \le OD_{patient} \le 1.2 \times OD_{cut-off}$

Positive OD patient > 1.2 x OD cut-off

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9. Technical Data

Sample material: serum

Sample volume: 10 μl of sample diluted 1:101 with 1x sample buffer

Total incubation time: 90 minutes at 20-32°C/68-89.6°F

Calibration range: 0-300 U/ml

Analytical sensitivity: 1.0 U/ml

Storage: at 2-8°C/35-46°F use original vials, only

Number of determinations: 96 tests

10. Performance Data

10.1 Analytical sensitivity

Testing sample buffer 30 times on *AESKULISA Crohn's Check* gave an analytical sensivity of 1.0 U/ml.

10.2 Specificity and sensitivity

The microplate is coated with highly purified *mannan from S. cerevisiae*. No crossreactivities to other autoantigens have been found. The diagnostic specificity of the test is 90% for Crohn's disease. The diagnostic sensitivity is about 68%.

10.3 Linearity

Chosen sera have been tested with this kit and found to dilute linearly. However, due to the heterogeneous nature of human autoantibodies there might be samples that do not follow this rule.

		measured	expected	
Sample	Dilution	concentration	concentration	Recovery
No.	Factor	(U/ml)	(U/ml)	(%)
1	1 / 100	48.7	49.0	99.4
	1 / 200	23.8	24.5	97.1
	1 / 400	11.4	12.3	92.7
	1 / 800	5.9	6.1	96.7
2	1 / 100	28.6	28.0	102.1
	1 / 200	13.8	14.0	98.6
	1 / 400	7.0	7.0	100.0
	1 / 800	3.4	3.5	97.1

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10.4 Precision

To determine the precision of the assay, the variability (intra and inter-assay) was assessed by examining its reproducibility on three serum samples selected to represent a range over the standard curve.

Intra-Assay								
Sample Mean CV								
No.	(U/ml)	(%)						
1	2.7	0.3						
2	23.7	2.7						
3	46.0	3.0						

Inter-Assay								
Sample Mean CV								
No.	(U/ml)	(%)						
1	2.3	0.9						
2	27.3	3.9						
3	54.3	5.7						

10.5 Calibration

Due to the lack of international reference calibration this assay is calibrated in arbitrary units (U/ml).

11. Literature

1. Seibold F, Stich O, Hufnagel R, Kamil S, Scheurlen M (2001).

Anti-Saccharomyces cerevisiae antibodies in inflammatory bowel disease: a family study. Scand J Gastroenterol 36: 196-201.

2. Sendid B, Colombel JF, Jacquinot PM et al.(1996).

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3. Giaffer MH, Clark A, Holdsworth CD (1992).

Antibodies to Saccharomyces cerevisiae in patients with Crohn's disease and their possible pathogenic importance.

Gut 33: 1071-1075.

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ANNEX A: Pipetting scheme

We suggest pipetting calibrators, controls and samples as follows:

For quantitative interpretation use calibrators to establish a standard curve.

For qualitative interpretation use cut-off calibrator.

	for quantitative interpretation use calibrators to establish a standard curve					li- for qualitative interpretation use cut- off calibrator				cut-		
	1	2	3	4	5	6	7	8	9	10	11	12
Α	CalA	CalE	P1				NC	P2				
В	CalA	CalE	P1				NC	P2				
С	CalB	CalF	P2				CC	P3				
D	CalB	CalF	P2				CC	P3				
E	CalC	PC	P3				PC					
F	CalC	PC	P3				PC					
G	CalD	NC					P1					
Н	CalD	NC					P1					

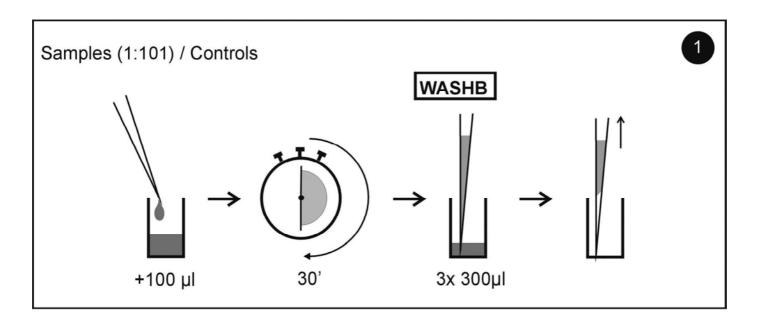
CalA: calibrator A, CalB: calibrator B, CalC: calibrator C, CalD: calibrator D, CalE: calibrator E,

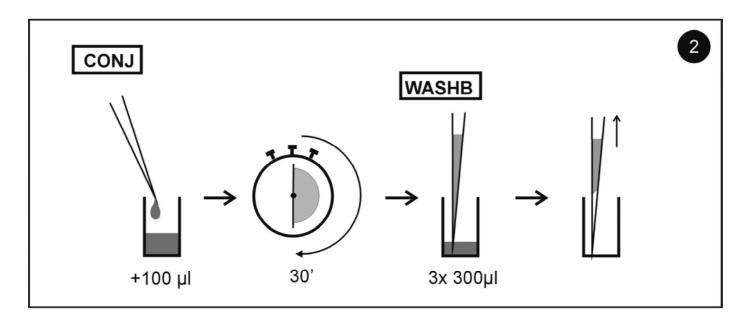
CalF: calibrator F PC: positive control NC: negative control CC: Cut-off calibrator

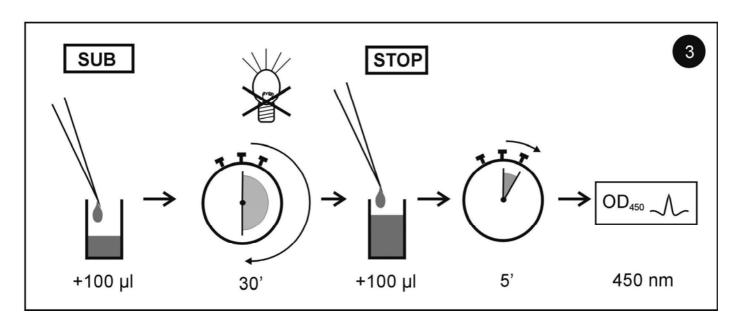
P1: patient 1 P2: patient 2 P3: patient 3

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Annex B: Test Procedure







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AESKU.DIAGNOSTICS GmbH 55234 Wendelsheim - Mikroforum Ring 2, Germany Phone: + 49-6734-96270, Fax: + 49-6734-962727

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	◆ Déclaração CE de Conformidade	
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	◆ 96 Bestimmungen	◆ 96 προσδιορισμοί
	♦ 96 Testes	
	♦ Rispettare le istruzioni per l'uso	 See instructions for use
	♦ Voir les instructions d'utilisation	 Ver las instrucciones de uso
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WASHB 50x	◆ Tampon de Lavage	♦ Solución de lavado
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	♦ Solucão de lavagem	
	Tampone substrato	Substrate buffer
SUB	♦ Substrat	♦ Tampón sustrato
308	Substratpuffer	 Ρυθμιστικό διάλυμα υποστρώματος
	◆ Substrato	
	♦ Reagente bloccante	◆ Stop solution
CTOD	♦ Solution d'Arrêt	♦ Solución de parada
[3107]	◆ Stopreagenz	 Αντιδραστήριο διακοπής αντίδρασης
	♦ Solução de paragem	
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SB 5x	♦ Probenpuffer	 Ρυθμιστικό διάλυμα δειγμάτων
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