

AESKULISA HiT II Check

REF 3291

Instruction manual

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1. Intended Use

AESKULISA HiT II Check is a solid phase enzyme immunoassay for the quantitative and qualitative detection of antibodies that cause heparin induced thrombocytopenia type II.

2. Clinical Application and Principle of the Assay

Heparin induced thrombocytopenia (HiT) is a severe side-effect of heparin treatment and occurs in 1-3 % of the treated patients.

Two distinct types of heparin induced thrombocytopenia can occur: HiT type I has no clinical relevance and is characterised by a transient decrease in platelet counts. It recovers after a few days even if the administration of heparin continues. HiT type II is an immune-mediated form. The platelet counts drop more than 50 % from the baseline 5 to 14 days after first beginning of heparin administration. The affected patients develop antibodies that recognize neoepitopes exposed by the complex of platelet factor 4 and heparin. The antibodies found are most commonly of the IgG subclass with or without IgM and IgA class antibodies. IgM and IgA are rarely found without IgG antibodies. However, the pathogenic effect has been proofed for the IgG subclass whereas the effect of IgM and IgA antibodies is still controversially discussed. The Fc portion of the HiT antibody can bind to the platelet Fc receptor and this interaction triggers activation and aggregation of the platelets. Activated platelets release PF4, thus perpetuating the cycle of heparin-induced platelet activation. The platelet activation leads to the production of platelet factor 4 and prothrombotic platelet microparticles. Heparin-like molecules (heparan sulfate) on the surface of endothelial cells are also able to form a complex with platelet factor 4 that can be recognized by HiT antibodies. That, in turn, can induce tissue factor expression with further activation of the coagulation cascade and thrombin generation. It leads to an increased risk for new arterial and venous thromboembolic complications that can be lethal in 10-15 % of the patients.

An early diagnosis and a replacement with a suitable alternative anticoagulant can clearly minimize the complication rate.

Principle of the test

Plasma samples diluted 1:101 are incubated in the microplates coated with the specific antigen. Patient's antibodies, if present in the specimen, bind to the antigen. The unbound fraction is washed off in the following step. Afterwards anti-human immunoglobulins conjugated to horseradish peroxidase (conjugate) are incubated and react with the antigen-antibody complex of the samples in the microplates. Unbound conjugate is washed off in the following step. Addition of TMB-substrate generates an enzymatic colorimetric (blue) reaction, which is stopped by diluted acid (color changes to yellow). The rate of color formation from the chromogen is a function of the amount of conjugate bound to the antigen-antibody complex and this is proportional to the initial concentration of the respective antibodies in the patient sample.

3. Kit Contents

To be reconstituted:

5x Sample Buffer 1 vial, 20 ml - 5x concentrated (capped white: yellow solution)

Containing: Tris, NaCl, BSA, sodium azide < 0.1% (preservative)

50x Wash Buffer 1 vial, 20 ml - 50x concentrated (capped white: green solution)

Containing: Tris, NaCl, Tween 20, sodium azide < 0.1% (preservative)

Ready to use:

Negative Control 1 vial, 1.5 ml (capped green: colorless solution)

Containing: Human plasma (diluted), sodium azide < 0.1% (preservative)

Positive Control 1 vial, 1.5 ml (capped red: yellow solution)

Containing: Human plasma (diluted), sodium azide < 0.1% (preservative)

Cut-off Calibrator 1 vial, 1.5 ml (capped blue: yellow solution)

Containing: Human plasma (diluted), sodium azide < 0.1% (preservative)

Calibrators 6 vials, 1.5 ml each 0, 3, 10, 30, 100, 300 U/ml

(color increasing with concentration: yellow solutions)

Containing: Human plasma (diluted), sodium azide < 0.1% (preservative)

Conjugates 1 vial, 15 ml IgA/G/M (capped white: red solution)

Containing: Anti-human immunoglobulins conjugated to horseradish peroxidase

TMB Substrate 1 vial, 15 ml (capped black)

Containing: Stabilized TMB/H₂O₂

Stop Solution 1 vial, 15 ml (capped white: colorless solution)

Containing: 1M Hydrochloric Acid

Microtiter plate 12x8 well strips with breakaway microwells

Coating see paragraph 1

Material required but not provided:

Microtiter plate reader 450 nm reading filter and optional 620 nm reference filter (600-690 nm). Glass ware (cylinder 100-1000ml), test tubes for dilutions. Vortex mixer, precision pipettes (10, 100, 200, 500, 1000 µl) or adjustable multipipette (100-1000 ml). Microplate washing device (300 µl repeating or multichannel pipette or automated system), adsorbent paper.

Our tests are designed to be used with purified water according to the definition of the United States Pharmacopeia (USP 26 - NF 21) and the European Pharmacopeia (Eur.Ph. 4th ed.).

4. Storage and Shelf Life

Store all reagents and the microplate at 2-8°C/35-46°F, in their original containers. Once prepared, reconstituted solutions are stable for 1 month at 4°C/39°F, at least. **Reagents and the microplate shall be used within the expiry date indicated on each component, only. Avoid intense exposure of TMB solution to light. Store microplates in designated foil, including the desiccant, and seal tightly.**

5. Precautions of Use

5.1 Health hazard data

THIS PRODUCT IS FOR IN VITRO DIAGNOSTIC USE ONLY. Thus, only staff trained and specially advised in methods of in vitro diagnostics may perform the kit. Although this product is not considered particularly toxic or dangerous in conditions of normal use, refer to the following for maximum safety :

Recommendations and precautions

This kit contains potentially hazardous components. Though kit reagents are not classified being irritant to eyes and skin we recommend to avoid contact with eyes and skin and wear disposable gloves.

WARNING ! Calibrators, Controls and Buffers contain sodium azide (NaN_3) as a preservative. NaN_3 may be toxic if ingested or adsorbed by skin or eyes. NaN_3 may react with lead and copper plumbing to form highly explosive metal azides. On disposal, flush with a large volume of water to prevent azide build-up. Please refer to decontamination procedures as outlined by CDC or other local/national guidelines.

Do not smoke, eat or drink when manipulating the kit.

Do not pipette by mouth.

All human source material used for some reagents of this kit (controls, standards e.g.) has been tested by approved methods and found negative for HbsAg, Hepatitis C and HIV 1. However, no test can guarantee the absence of viral agents in such material completely. Thus handle kit controls, standards and patient samples as if capable of transmitting infectious diseases and according to national requirements.

5.2 General directions for use

Do not mix or substitute reagents or microplates from different lot numbers. This may lead to variations in the results.

Allow all components to reach room temperature (20-32 °C/68-89.6 °F) before use, mix well and follow the recommended incubation scheme for an optimum performance of the test.

Incubation: We recommend test performance at 30 °C/86 °F for automated systems.

Never expose components to higher temperature than 37 °C/ 98.6 °F.

Always pipette substrate solution with brand new tips only. Protect this reagent from light. Never pipette conjugate with tips used with other reagents prior.

A definite clinical diagnosis should not be based on the results of the performed test only, but should be made by the physician after all clinical and laboratory findings have been evaluated. The diagnosis is to be verified using different diagnostic methods.

6. Sample Collection, Handling and Storage

Use preferentially plasma samples freshly collected with 3.2% or 3.8% sodium citrate as an anticoagulant. Blood withdrawal must follow national requirements. Do not use icteric, lipemic, hemolysed or bacterially contaminated samples. Blood samples should be collected in clean, dry and empty tubes. After centrifugation, the plasma samples should be used immediately, otherwise stored tightly closed at 2-8 °C/35-46 °F up to eight hours, or frozen at -20 °C/-4 °F for longer periods.

Do not use samples anticoagulated with heparin in this assay!

7. Assay Procedure

7.1 Preparations prior to pipetting

Dilute concentrated reagents:

Dilute the concentrated sample buffer 1:5 with distilled water (e.g. 20 ml plus 80 ml).

Dilute the concentrated wash buffer 1:50 with distilled water (e.g. 20 ml plus 980 ml).

Samples:

Dilute plasma samples 1:101 with sample buffer (1x)

e.g. 1000 µl sample buffer (1x) + 10 µl plasma. Mix well !

Washing:

Prepare 20 ml of diluted wash buffer (1x) per 8 wells or 200 ml for 96 wells

e.g. 4 ml concentrate plus 196 ml distilled water.

Automated washing:

Consider excess volumes required for setting up the instrument and dead volume of robot pipette.

Manual washing:

Discard liquid from wells by inverting the plate. Knock the microwell frame with wells downside vigorously on clean adsorbent paper. Pipette 300 µl of diluted wash buffer into each well, wait for 20 seconds. Repeat the whole procedure twice again.

Microplates:

Calculate the number of wells required for the test. Remove unused wells from the frame, replace and store in the provided plastic bag, together with desiccant, seal tightly (2-8 °C/35-46 °F).

7.2 Work flow

For pipetting scheme see Annex A, for the test procedure see Annex B

We recommend pipetting samples and calibrators in duplicate.

Cut-off calibrator should be used for qualitative testing only.

- Pipette 100 µl of each patient's diluted plasma into the designated microwells.
- Pipette 100 µl calibrators OR cut-off calibrator and negative and positive controls into the designated wells.
- Incubate for 30 minutes at 20-32 °C/68-89.6 °F.
- Wash 3x with 300 µl washing buffer (diluted 1:50).
- Pipette 100 µl conjugate into each well.
- Incubate for 30 minutes at 20-32 °C/68-89.6 °F.
- Wash 3x with 300 µl washing buffer (diluted 1:50).
- Pipette 100 µl TMB substrate into each well.
- Incubate for 30 minutes at 20-32 °C/68-89.6 °F, protected from intense light.
- Pipette 100 µl stop solution into each well, using the same order as pipetting the substrate.
- Incubate 5 minutes minimum.
- Agitate plate carefully for 5 sec.
- Read absorbance at 450 nm (optionally 450/620 nm) within 30 minutes.

8. Quantitative and Qualitative Interpretation

For **quantitative interpretation** establish the standard curve by plotting the **optical density (OD) of each calibrator (y-axis)** with respect to the corresponding concentration values in **U/ml (x-axis)**. For best results we recommend log/lin coordinates and 4-Parameter Fit. From the OD of each sample, read the corresponding antibody concentrations expressed in **U/ml**.

Normal Range	Equivocal Range	Positive Results
< 16 U/ml	16 - 24 U/ml	> 24 U/ml

Example of a standard curve

We recommend pipetting calibrators in parallel for each run.

Calibrators IgA/G/M	OD 450/620 nm	CV % (Variation)
0 U/ml	0.025	0.0
3 U/ml	0.139	3.5
10 U/ml	0.283	4.3
30 U/ml	0.598	4.0
100 U/ml	1.224	3.6
300 U/ml	2.123	2.8

Example of calculation

Patient	Replicate (OD)	Mean (OD)	Result (U/ml)
P 01	0.793/0.801	0.797	47.7
P 02	0.308/0.333	0.321	12.1

For lot specific data, see enclosed quality control leaflet. Medical laboratories might perform an in-house Quality Control by using own controls and/or internal pooled plasma, as foreseen by EU regulations.

Do not use this example for interpreting patients results!

Each laboratory should establish its own normal range based upon its own techniques, controls, equipment and patient population according to their own established procedures.

For qualitative interpretation read the optical density of the cut-off calibrator and the patient samples. Compare patient's OD with the OD of the cut-off calibrator. For qualitative interpretation we recommend to consider plasma within a range of 20% around the cut-off value as equivocal. All samples with higher ODs are considered positive, samples with lower ODs are considered negative.

Negative: $OD_{\text{patient}} < 0.8 \times OD_{\text{cut-off}}$

Equivocal: $0.8 \times OD_{\text{cut-off}} \leq OD_{\text{patient}} \leq 1.2 \times OD_{\text{cut-off}}$

Positive $OD_{\text{patient}} > 1.2 \times OD_{\text{cut-off}}$

9. Technical Data

Sample material:	plasma
Sample volume:	10 µl of sample diluted 1:101 with 1x sample buffer
Total incubation time:	90 minutes at 20-32 °C/68-89.6 °F
Calibration range:	0-300 U/ml
Analytical sensitivity:	1.0 U/ml
Storage:	at 2-8 °C/35-46 °F use original vials, only
Number of determinations:	96 tests

10. Performance Data

10.1 Analytical sensitivity

Testing sample buffer 30 times on *AESKULISA HiT II Check* gave an analytical sensitivity of 1.0 U/ml.

10.2 Specificity and sensitivity

The sensitivity of the *AESKULISA HiT II Check* was determined to be 100 % in comparison to plasma samples with known immune status. Clinically defined plasma samples show a specificity of 90%.

10.3 Linearity

Chosen plasma samples have been tested with this kit and found to dilute linearly. However, due to the heterogeneous nature of human autoantibodies there might be samples that do not follow this rule.

Sample No.	Dilution Factor	measured concentration (U/ml)	expected concentration (U/ml)	Recovery (%)
1	1 / 100	77.6	80.0	97.0
	1 / 200	37.5	40.0	93.8
	1 / 400	18.8	20.0	94.0
	1 / 800	9.1	10.0	91.0
2	1 / 100	7.7	8.0	96.3
	1 / 200	3.8	4.0	95.0
	1 / 400	2.2	2.0	110.0
	1 / 800	1.0	1.0	100.0

10.4 Precision

To determine the precision of the assay, the variability (intra and inter-assay) was assessed by examining its reproducibility on three plasma samples selected to represent a range over the standard curve.

Intra-Assay		
Sample No.	Mean (U/ml)	CV (%)
1	9.8	1.3
2	120.2	9.8
3	210.2	10.0

Inter-Assay		
Sample No.	Mean (U/ml)	CV (%)
1	10.2	1.0
2	106.6	9.5
3	208.6	6.3

10.5 Calibration

Due to the lack of international reference calibration this assay is calibrated in arbitrary units (U/ml).

11. Literature

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New approaches to the diagnosis of heparin induced thrombocytopenia.
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- 2. Franchini M. (2005).**
Heparin induced thrombocytopenia: an update.
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- 3. Warkentin T.E. (2004).**
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- 4. Lindhoff-Last E., Gerdson F., Ackermann H., Bauersachs R. (2001).**
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- 5. Warkentin T.E., Chong B.H., Greinacher A. (1998).**
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- 7. Greinacher A., Poetzsch B., Amiral J., Dummel V., Eichner A., Mueller-Eckhardt C. (1994).**
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Thrombosis and Haemostasis 71: 247-251.
- 8. Chong B.H., Fawaz I., Chesterman C.N., Berndt M.C. (1989).**
Heparin induced thrombocytopenia: mechanism of interaction of the heparin-dependent antibody with platelets.
British Journal of Haematology 73: 235-240.

ANNEX A: Pipetting scheme

We suggest pipetting calibrators, controls and samples as follows:

For **quantitative interpretation** use calibrators to establish a standard curve.

For **qualitative interpretation** use cut-off calibrator.

	for quantitative interpretation use calibrators to establish a standard curve						for qualitative interpretation use cut-off control					
	1	2	3	4	5	6	7	8	9	10	11	12
A	CalA	CalE	P1				NC	P2				
B	CalA	CalE	P1				NC	P2				
C	CalB	CalF	P2				CC	P3				
D	CalB	CalF	P2				CC	P3				
E	CalC	PC	P3				PC	...				
F	CalC	PC	P3				PC	...				
G	CalD	NC	...				P1	...				
H	CalD	NC	...				P1	...				

CalA: calibrator A, CalB: calibrator B, CalC: calibrator C, CalD: calibrator D, CalE: calibrator E, CalF: calibrator F

PC: positive control

NC: negative control

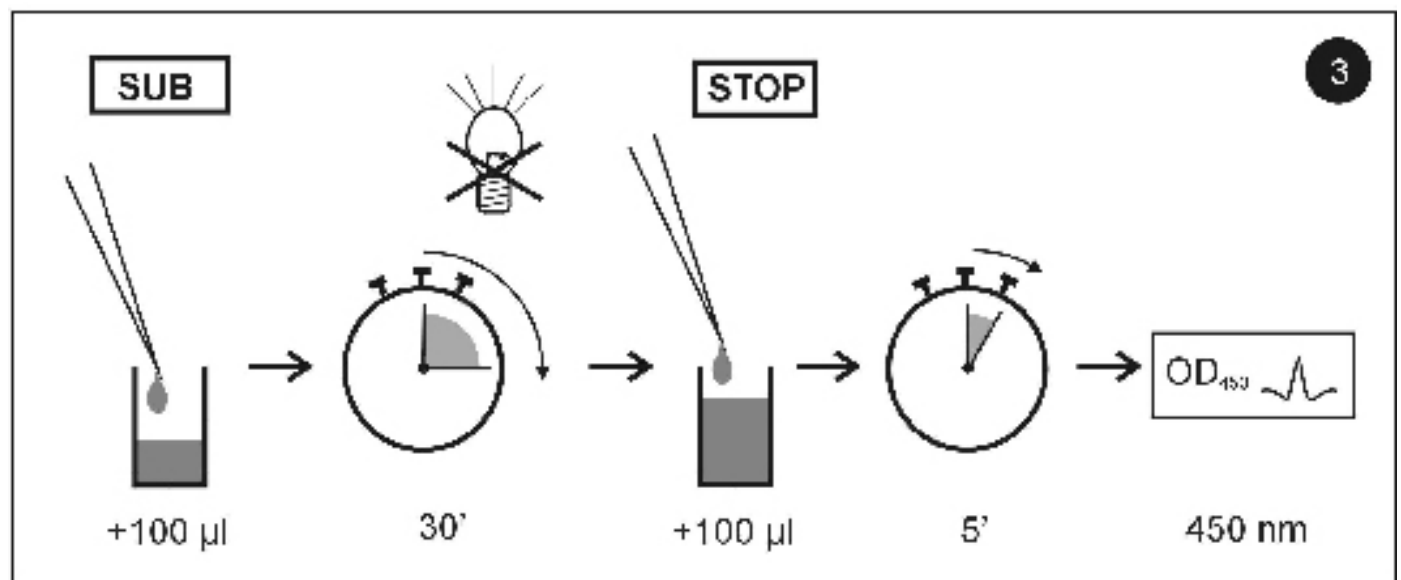
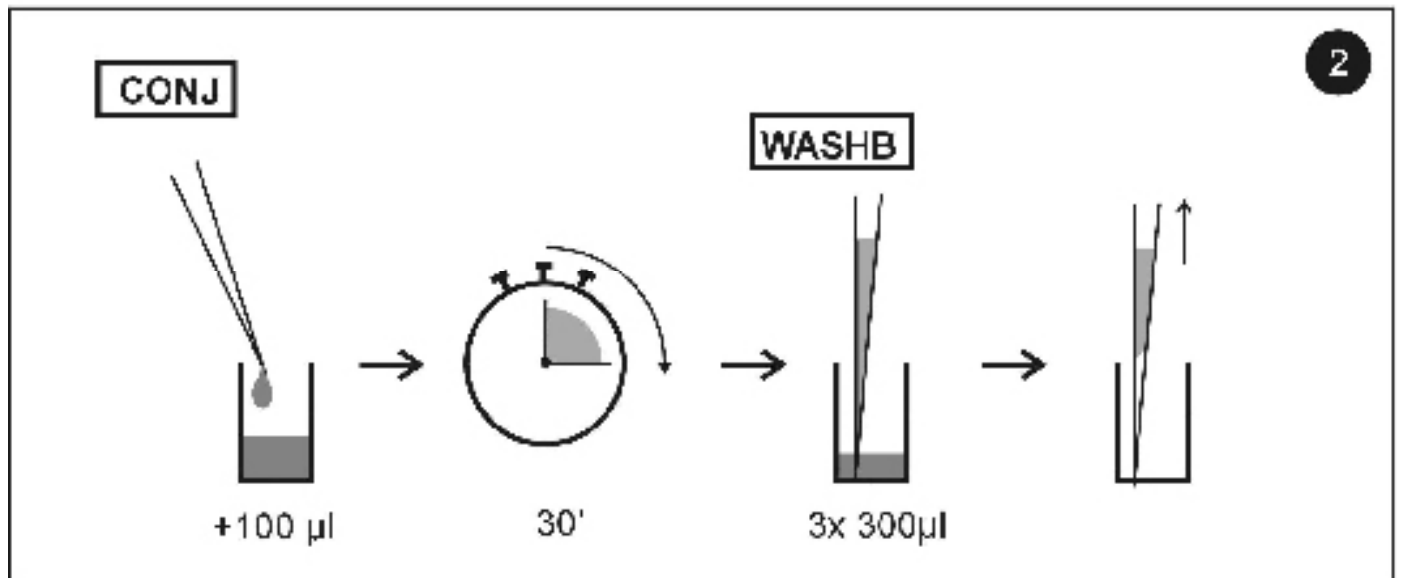
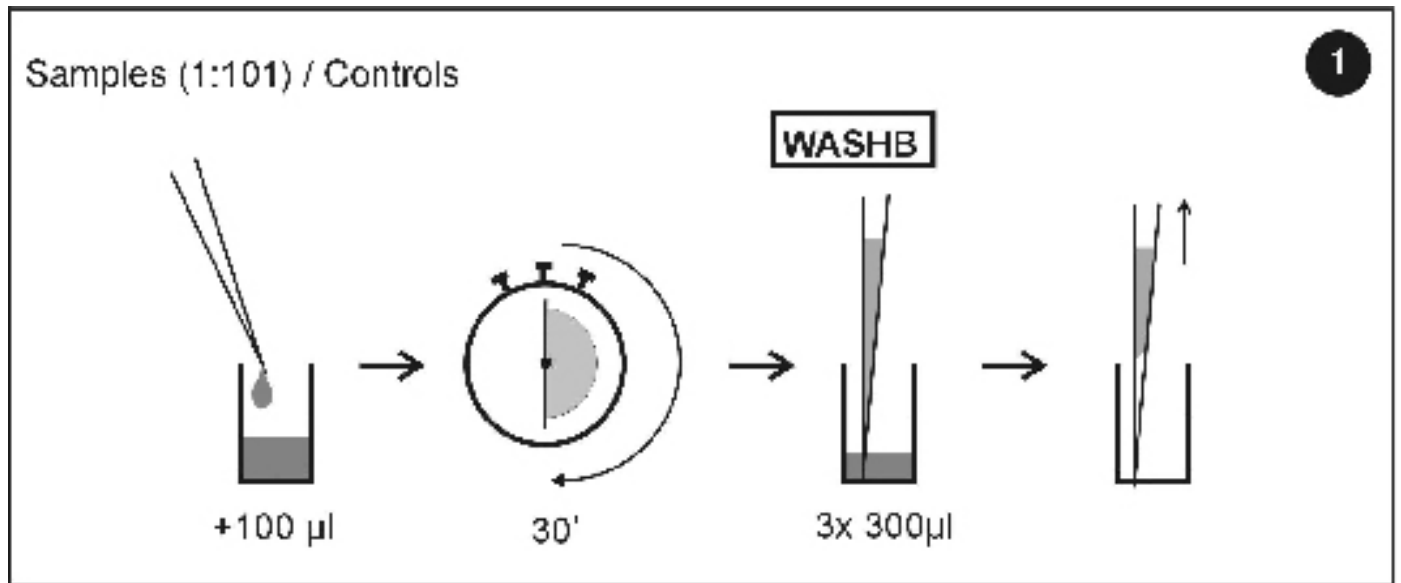
CC: Cut-off calibrator

P1: patient 1

P2: patient 2

P3: patient 3










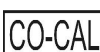




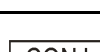
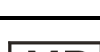
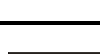
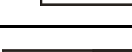
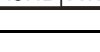
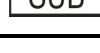
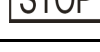
Annex B: Test Procedure



Assay/Test: _____ Incubation / Inkub. : 1. _____ min Date/Datum: _____

Temperature/Temperatur: _____ °F _____ °C 2. _____ min
Signature/Unterschrift: _____
Name: _____ 3. _____ min

	1	2	3	4	5	6	7	8	9	10	11	12
A												
B												
C												
D												
E												
F												
G												
H												

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	<ul style="list-style-type: none"> ◆ Da utilizzarsi entro ◆ Utilise avant le ◆ Verwendbar bis ◆ Utilizar antes de 	<ul style="list-style-type: none"> ◆ Use by ◆ Utilizar antes de ◆ Χρήση μέχρι
	<ul style="list-style-type: none"> ◆ Conservare a 2-8°C ◆ Conserver à 2-8°C ◆ Lagerung bei 2-8°C ◆ Conservar entre 2-8°C 	<ul style="list-style-type: none"> ◆ Store at 2-8°C (35-46°F) ◆ Conservar a 2-8°C ◆ Φυλάσσεται στους 2-8°C
	<ul style="list-style-type: none"> ◆ Prodotto da ◆ Fabriqué par ◆ Hergestellt von ◆ Fabricado por 	<ul style="list-style-type: none"> ◆ Manufactured by ◆ Fabricado por ◆ Κατασκευάζεται από
	<ul style="list-style-type: none"> ◆ Calibratore cut-off ◆ Etalon Seuil ◆ Grenzwert Kalibrator ◆ Calibrador de cut-off 	<ul style="list-style-type: none"> ◆ Cut off Calibrator ◆ Calibrador de cut-off ◆ Οριακός ορός Αντιδραστήριο βαθμονόμησης
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	<ul style="list-style-type: none"> ◆ Tampone di lavaggio ◆ Tampon de Lavage ◆ Waschpuffer ◆ Solução de lavagem 	<ul style="list-style-type: none"> ◆ Wash buffer ◆ Solución de lavado ◆ Ρυθμιστικό διάλυμα πλύσης
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