AESKULISA Nucleo-h

REF 3130

Instruction manual

Contents

1. Intended Use	1
2. Clinical Applications and Principle of the Assay	1
3. Kit Contents	2
4. Storage and Shelf Life	2
5. Precautions of Use	3
6. Sample Collection, Handling and Storage	3
7. Assay Procedure	4
8. Quantitative and Qualitative Interpretation	5
9. Technical Data	6
10. Performance Data	6-7
11. Literature	7
A : Pipetting scheme	8
B : Test Procedure	9

1. Intended Use

AESKULISA Nucleo-h is a solid phase enzyme immunoassay with human native nucleosomes isolated from the eukaryotic celline HeLa for the quantitative and qualitative detection of antibodies against nucleosomes and their components dsDNA and histones in human serum.

The assay is a tool in the differential diagnosis of systemic lupus erythematosus (SLE).

2. Clinical Application and Principle of the Assay

Nucleosomes are the basic structure of chromatin important for the compaction of DNA in the nucleus. A nucleosome comprises of four histones H2A, H2B, H3 and H4 forming an octamer and 146 bp of dsDNA wrapped 1 ³/₄ turns around the core (H2A, H2B, H3, H4)₂ octamer. Histone H1 interacts with the nucleosome and together with linked-DNA connects neighboring nucleosomes.

Antibodies against double-stranded DNA (dsDNA) and against histones are common features of systemic lupus erythematosus (SLE). Burlingame at al. (1993) were the first to show in murine models that the production of anti-chromatin antibodies is induced by a T-cell dependent immune response with self antigen. It has been demonstrated that the initial autoimmune response in murine models of SLE was directed against the nucleosome. Thus anti-nucleosome specific antibodies represent specific and early markers of SLE recognizing conformational epitopes shared by the native nucleosome molecule. Later the autoimmune response can diversify to the components of the nucleosomes, DNA and histones, as a part of intermolecular spreading.

Nucleosome-specific antibodies are found in 84-88 % of SLE patients.

Principle of the test

Serum samples diluted 1:101 are incubated in the microplates coated with the specific antigen. Patient's antibodies, if present in the specimen, bind to the antigen. The unbound fraction is washed off in the following step. Afterwards anti-human immunoglobulins conjugated to horseradish peroxidase (conjugate) are incubated and react with the antigen-antibody complex of the samples in the microplates. Unbound conjugate is washed off in the following step. Addition of TMB-substrate generates an enzymatic colorimetric (blue) reaction, which is stopped by diluted acid (color changes to yellow). The rate of color formation from the chromogen is a function of the amount of conjugate bound to the antigen-antibody complex and this is proportional to the initial concentration of the respective antibodies in the patient sample.

3. Kit Contents

<i>To be reconstitute</i> 5x Sample Buffer	ed: 1 vial, 20 ml - 5x concentrated (capped white: yellow solution) Containing: Tris, NaCl, BSA, sodium azide < 0.1% (preservative)
50x Wash Buffer	1 vial, 20 ml - 50x concentrated (capped white: green solution) Containing: Tris, NaCl, Tween 20, sodium azide < 0.1% (preservative)
Ready to use: Negative Control	1 vial, 1.5 ml (capped green: colorless solution) Containing: Human serum (diluted), sodium azide < 0.1% (preservative)
Positive Control	1 vial, 1.5 ml (capped red: yellow solution) Containing: Human serum (diluted), sodium azide < 0.1% (preservative)
Cut-off Calibrator	1 vial, 1.5 ml (capped blue: yellow solution) Containing: Human serum (diluted), sodium azide < 0.1% (preservative)
Calibrators	6 vials, 1.5 ml each 0, 3, 10, 30, 100, 300 U/ml (color increasing with concentration: yellow solutions) Containing: Human serum (diluted), sodium azide < 0.1% (preservative)
Conjugate	1 vial,15 ml IgG (capped blue: blue solution) Containing: Anti-human immunoglobulins conjugated to horseradish peroxidase
TMB Substrate	1 vial, 15 ml (capped black) Containing: Stabilized TMB/H2O2
Stop Solution	1 vial, 15 ml (capped white: colorless solution) Containing: 1M Hydrochloric Acid
Microtiterplate	12x8 well strips with breakaway microwells Coating see paragraph 1

Material required but not provided:

Microtiter plate reader 450 nm reading filter and optional 620 nm reference filter (600-690 nm). Glass ware(cylinder 100-1000ml), test tubes for dilutions. Vortex mixer, precision pipettes (10, 100, 200, 500, 1000 μ l) or adjustable multipipette (100-1000ml). Microplate washing device (300 μ l repeating or multi-channel pipette or automated system), adsorbent paper.

Our tests are designed to be used with purified water according to the definition of the United States Pharmacopeia (USP 26 - NF 21) and the European Pharmacopeia (Eur.Ph. 4th ed.).

4. Storage and Shelf Life

Store all reagents and the microplate at 2-8°C/35-46°F, in their original containers. Once prepared, reconstituted solutions are stable for 1 month at 4°C/39°F, at least. *Reagents and the microplate shall be used within the expiry date indicated on each component, only. Avoid intense exposure of TMB solution to light. Store microplates in designated foil, including the desiccant, and seal tightly.*

5.1 Health hazard data

THIS PRODUCT IS FOR IN VITRO DIAGNOSTIC USE ONLY. Thus, only staff trained and specially advised in methods of in vitro diagnostics may perform the kit. Although this product is not considered particularly toxic or dangerous in conditions of normal use, refer to the following for maximum safety :

Recommendations and precautions

This kit contains potentially hazardous components. Though kit reagents are not classified being irritant to eyes and skin we recommend to avoid contact with eyes and skin and wear disposable gloves.

WARNING ! Calibrators, Controls and Buffers contain sodium azide (NaN_3) as a preservative. NaN_3 may be toxic if ingested or adsorbed by skin or eyes. NaN_3 may react with lead and copper plumbing to form highly explosive metal azides. On disposal, flush with a large volume of water to prevent azide build-up. Please refer to decontamination procedures as outlined by CDC or other local/national guidelines.

Do not smoke, eat or drink when manipulating the kit.

Do not pipette by mouth.

All human source material used for some reagents of this kit (controls, standards e.g.) has been tested by approved methods and found negative for HbsAg, Hepatitis C and HIV 1. However, no test can guarantee the absence of viral agents in such material completely. Thus handle kit controls, standards and patient samples as if capable of transmitting infectious diseases and according to national requirements.

5.2 General directions for use

Do not mix or substitute reagents or microplates from different lot numbers. This may lead to variations in the results.

Allow all components to reach room temperature (20-32°C/68-89.6°F) before use, mix well and follow the recommended incubation scheme for an optimum performance of the test.

Incubation: We recommend test performance at 30°C/86°F for automated systems.

Never expose components to higher temperature than 37°C/ 98.6 °F.

Always pipette substrate solution with brand new tips only. Protect this reagent from light. Never pipette conjugate with tips used with other reagents prior.

A definite clinical diagnosis should not be based on the results of the performed test only, but should be made by the physician after all clinical and laboratory findings have been evaluated. The diagnosis is to be verified using different diagnostic methods.

6. Sample Collection, Handling and Storage

Use preferentially freshly collected serum samples. Blood withdrawal must follow national requirements.

Do not use icteric, lipemic, hemolysed or bacterially contaminated samples. Sera with particles should be cleared by low speed centrifugation (<1000 x g). Blood samples should be collected in clean, dry and empty tubes. After separation, the serum samples should be used immediately, respectively stored tightly closed at 2-8°C/35-46°F up to three days, or frozen at -20°C/-4°F for longer periods.

7.1 Preparations prior to pipetting

Dilute concentrated reagents:

Dilute the concentrated sample buffer 1:5 with distilled water (e.g. 20 ml plus 80 ml). Dilute the concentrated wash buffer 1:50 with distilled water (e.g. 20 ml plus 980 ml).

Samples:

Dilute serum samples 1:101 with sample buffer (1x) e.g. 1000 μ l sample buffer (1x) + 10 μ l serum. Mix well !

Washing:

Prepare 20 ml of diluted wash buffer (1x) per 8 wells or 200 ml for 96 wells e.g. 4 ml concentrate plus 196 ml distilled water.

Automated washing:

Consider excess volumes required for setting up the instrument and dead volume of robot pipette.

Manual washing:

Discard liquid from wells by inverting the plate. Knock the microwell frame with wells downside vigorously on clean adsorbent paper. Pipette 300 μ l of diluted wash buffer into each well, wait for 20 seconds. Repeat the whole procedure twice again.

Microplates:

Calculate the number of wells required for the test. Remove unused wells from the frame, replace and store in the provided plastic bag, together with desiccant, seal tightly (2-8°C/35-46°F).

7.2 Work flow

For pipetting scheme see Annex A, for the test procedure see Annex B We recommend pipetting samples and calibrators in duplicate. Cut-off calibrator should be used for qualitative testing only.

- Pipette 100 µl of each patient's diluted serum into the designated microwells.
- Pipette 100 µl calibrators OR cut-off calibrator and negative and positive controls into the designated wells.
- Incubate for 30 minutes at 20-32°C/68-89.6°F.
- Wash 3x with 300 µl washing buffer (diluted 1:50).
- Pipette 100 µl conjugate into each well.
- Incubate for 30 minutes at 20-32°C/68-89.6°F.
- Wash 3x with 300 µl washing buffer (diluted 1:50).
- Pipette 100 µl TMB substrate into each well.
- Incubate for 30 minutes at 20-32°C/68-89.6°F, protected from intense light.
- Pipette 100 µl stop solution into each well, using the same order as pipetting the substrate.
- Incubate 5 minutes minimum.
- Agitate plate carefully for 5 sec.
- Read absorbance at 450 nm (optionally 450/620 nm) within 30 minutes.

8. Quantitative and Qualitative Interpretation

For **quantitative Interpretation** establish the standard curve by plotting the **optical density (OD) of each calibrator (y-axis)** with respect to the corresponding concentration values in **U/ml (x-axis)**. For best results we recommend log/lin coordinates and 4-Parameter Fit. From the OD of each sample, read the corresponding antibody concentrations expressed in **U/ml**.

Normal Range	Equivocal Range	Positive Results
< 12 U/ml	12 - 18 U/ml	>18 U/ml

Example of a standard curve

We recommend pipetting calibrators in parallel for each run.

Calibrators IgG	OD 450/620 nm	CV % (Variation)
0 U/ml	0.030	2.8
3 U/ml	0.136	1.0
10 U/ml	0.339	0.5
30 U/ml	0.661	1.4
100 U/ml	1.255	2.9
300 U/ml	2.131	1.8

Example of calculation

Patient	Replicate (OD)	Mean (OD)	Result (U/ml)
P 01	0.838/0.849	0.844	47.2
P 02	1.503/1.516	1.510	137.3

For lot specific data, see enclosed quality control leaflet. Medical laboratories might perform an inhouse Quality Control by using own controls and/or internal pooled sera, as foreseen by EU regulations.

Do not use this example for interpreting patients results!

Each laboratory should establish its own normal range based upon its own techniques, controls, equipment and patient population according to their own established procedures.

For qualitative interpretation read the optical density of the cut-off calibrator and the patient samples. Compare patient'sOD with the OD of the cut-off calibrator. For qualitative interpretation we recommend to consider sera within a range of 20% around the cut-off value as equivocal. All samples with higher ODs are considered positive, samples with lower ODs are considered negative.

Negative:	OD patient < 0.8 x OD cut-off
Equivocal:	$0.8 \times OD_{cut-off} \le OD_{patient} \le 1.2 \times OD_{cut-off}$
Positive	OD patient > 1.2 x OD cut-off

9. Technical Data

Sample material:	serum
Sample volume:	10 μ I of sample diluted 1:101 with 1x sample buffer
Total incubation time:	90 minutes at 20-32°C/68-89.6°F
Calibration range:	0-300 U/ml
Analytical sensitivity:	1.0 U/ml
Storage:	at 2-8°C/35-46°F use original vials, only
Number of determinations:	96 tests

10. Performance Data

10.1 Analytical sensitivity

Testing sample buffer 30 times on AESKULISA Nucleo-h gave an analytical sensivity of 1.0 U/ml.

10.2 Specificity and sensitivity

The microplates are coated with highly purified **native human nucleosomes.** No crossreactivities to other autoantigens have been found. The *AESKULISA* Nucleo-h Test shows a diagnostic specificity of 90%. The diagnostic sensitivity of nucleosome specific antibodies is 84-88% in SLE patients.

10.3 Linearity

Chosen sera have been tested with this kit and found to dilute linearly. However, due to the heterogeneous nature of human autoantibodies there might be samples that do not follow this rule.

		measured	expected	
Sample	Dilution	concentration	concentration	Recovery
No.	Factor	(U/ml)	(U/ml)	(%)
1	1 / 100	152.9	153.6	99.5
	1 / 200	76.4	76.8	99.4
	1 / 400	38.8	38.4	101.0
	1 / 800	17.8	19.2	92.7
2	1 / 100	85.3	84.9	100.5
	1 / 200	44.1	42.5	103.8
	1 / 400	22.0	21.2	103.8
	1 / 800	9.8	10.6	92.5

10.4 Precision

To determine the precision of the assay, the variability (intra and inter-assay) was assessed by examining its reproducibility on three serum samples selected to represent a range over the standard curve.

	Intra-Assa	ay
Sample	Mean	CV
No.	(U/ml)	(%)
1	174.02	3.7
2	77.98	2.1
3	32.57	2.3

	Inter-Assa	ay
Sample	Mean	CV
No.	(U/ml)	(%)
1	111.56	3.6
2	48.06	5.9
3	14.80	4.8

10.5 Calibration

Due to the lack of international reference calibration this assay is calibrated in arbitrary units (U/mI).

11. Literature

- Burlingame RW, Rubin RL, Balderas RS, Theofilopoulos AN (1993). Genesis and evolution of antichromatin autoantibodies in murine lupus indicates T-dependent immunization with self antigen. J Clin Invest 91: 1687-1696.
- Burlingame RW, Boey ML, Starkebaum G, Rubin RL (1994). The central role of chromatin in autoimmune response to histones and DNA in systemic lupus erythematosus. J Clin Invest 94: 184-192.
- **3.** Chabre H, Amoura Z, Piette JC, Godeau P, Bach JF, Koutouzov S (1995). *Presence of nucleosome-restricted antibodies in patients with systemic lupus erythematosus.* Arthritis Rheum 38: 1485-1491.

ANNEX A: Pipetting scheme

We suggest pipetting calibrators, controls and samples as follows: For quantitative interpretation use calibrators to establish a standard curve. For qualitative interpretation use cut-off calibrator.

	-			e rpreta a stand			-	alitativ ibrator	e inter	pretati	on use	cut-
	1	2	3	4	5	6	7	8	9	10	11	12
Α	CalA	CalE	P1				NC	P2				
В	CalA	CalE	P1				NC	P2				
С	CalB	CalF	P2				CC	P3				
D	CalB	CalF	P2				CC	P3				
Е	CalC	PC	P3				PC					
F	CalC	PC	P3				PC					
G	CalD	NC					P1					
Н	CalD	NC					P1					

CalA: calibrator A, CalB: calibrator B, CalC: calibrator C, CalD: calibrator D, CalE: calibrator E, CalF: calibrator F

PC: positive control

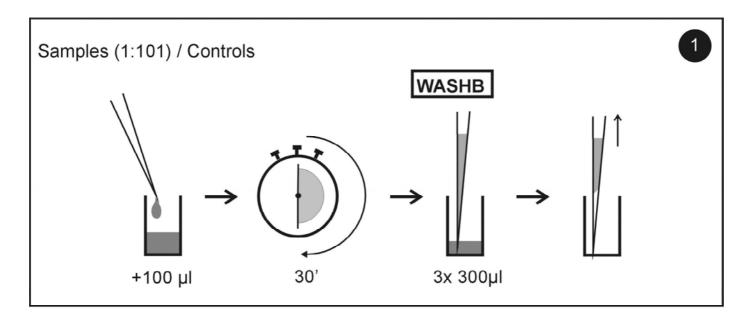
NC: negative control

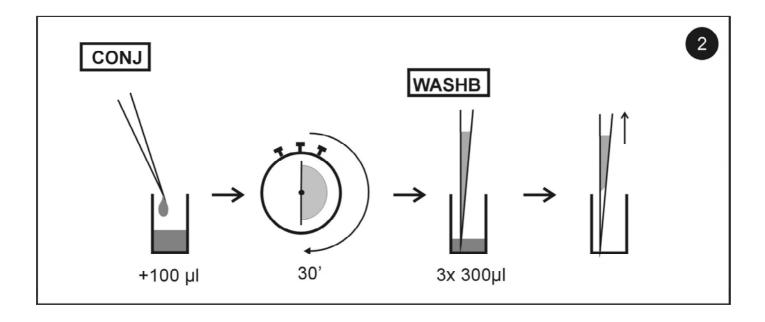
CC: Cut-off calibrator

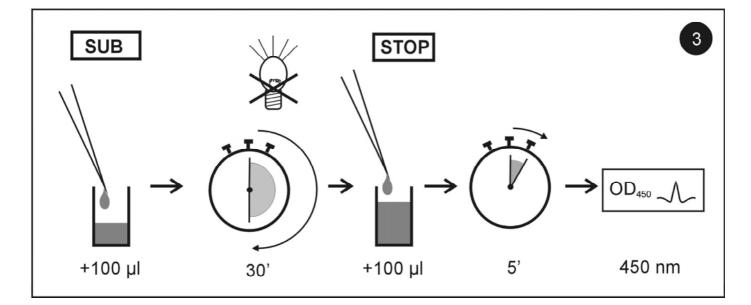
- P1: patient 1
- P2: patient 2

P3: patient 3

Annex B: Test Procedure







Assay/Test:			- -	Incubation / Inkub. :	IIIKUU.				Date	Date/ Datum:		
eratur	Temperature/Temperatur:	ur:	°F	°C		2.	min	U	. I / on the one i	torochrift.		
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	Diagnosi in vitro	 For in vitro diagnostic use
	Pour diagnostic in vitro	 Para uso diagnóstico in vitro
IVD	◆ In Vitro Diagnostikum	 In Vitro Διαγνωστικό μέσο
	 Para uso Diagnóstico in vitro 	
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	Bestellnummer	 Αριθμός παραγγελίας
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	◆ Lote	
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	♦ 96 Testes	
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-	♦ Prodotto da	 Manufactured by
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	♦ Hergestellt von	 Κατασκευάζεται από
	 Fabricado por 	
	 Calibratore cut-off 	 Cut off Calibrator
	 Etalon Seuil 	 Calibrador de cut-off
UU-UAL	 Grenzwert Kalibrator 	 Οριακός ορός Αντιδραστήριο βαθμονόμηση
	 Calibrador de cut-off 	
	 Controllo positivo 	 Positive Control
CON+	 Contrôle Positif 	 Control Positivo
	 Positiv Kontrolle 	 Θετικός ορός ελέγχου
	 Controlo positivo 	
	 Controllo negativo 	 Negative Control
CON	 Contrôle Négatif 	 Control Negativo
	 Negativ Kontrolle 	 Αρνητικός ορός ελέγχου
	 Controlo negativo 	
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CAL	♦ Etalon	♦ Calibrador
UAL	♦ Kalibrator	 Αντιδραστήριο βαθμονόμησης
	♦ Calibrador	
	♦ Recupero	♦ Recovery
RC	 Corrélation 	♦ Recuperado
	 Wiederfindung 	 Ανάκτηση
	♦ Recuperação	
	♦ Coniugato	♦ Conjugate
CONJ	♦ Conjugé	♦ Conjugado
00113	♦ Konjugat	 Σύζευγμα
	♦ Conjugado	
	 Micropiastra rivestita 	 Coated microtiter plate
MP	 Microplaque sensibilisée 	 Microplaca sensibilizada
	Beschichtete Mikrotiterplatte	 Επικαλυμμένη μικροπλάκα
	 Microplaca revestida 	
	 Piastra ad aghi rivestita 	 Coated pinplate
	 Pinplate sensibilisée 	 Pinplate sensibilizada
PINP	 Pinplate sensibilisée Beschichtete Pinplatte 	
PINP	 Pinplate sensibilisée 	 ♦ Pinplate sensibilizada ♦ Επικαλυμμένη πλάκα Pin
	 Pinplate sensibilisée Beschichtete Pinplatte Pinplate revestida Tampone di lavaggio 	 ◆ Pinplate sensibilizada ◆ Επικαλυμμένη πλάκα Pin ◆ Wash buffer
	 Pinplate sensibilisée Beschichtete Pinplatte Pinplate revestida Tampone di lavaggio Tampon de Lavage 	 Pinplate sensibilizada Επικαλυμμένη πλάκα Pin Wash buffer Solución de lavado
PINP WASHB 50x	 Pinplate sensibilisée Beschichtete Pinplatte Pinplate revestida Tampone di lavaggio Tampon de Lavage Waschpuffer 	 ◆ Pinplate sensibilizada ◆ Επικαλυμμένη πλάκα Pin ◆ Wash buffer
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	 Pinplate sensibilisée Beschichtete Pinplatte Pinplate revestida Tampone di lavaggio Tampon de Lavage Waschpuffer Solucão de lavagem Tampone substrato 	 Pinplate sensibilizada Επικαλυμμένη πλάκα Pin Wash buffer Solución de lavado Ρυθμιστικό διάλυμα πλύσης Substrate buffer
WASHB 50x	Pinplate sensibilisée Beschichtete Pinplatte Pinplate revestida Tampone di lavaggio Tampon de Lavage Waschpuffer Solucão de lavagem Tampone substrato Substrat	 Pinplate sensibilizada Επικαλυμμένη πλάκα Pin Wash buffer Solución de lavado Ρυθμιστικό διάλυμα πλύσης Substrate buffer Tampón sustrato
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