

MAGLUMI Rubella IgG (CLIA)



130212003M



100



**Shenzhen New Industries
Biomedical Engineering Co., Ltd**
4F, Wearnes Tech Bldg,
Science & Industry Park,
Nanshan, Shenzhen, 518057 CHINA
Tel. + 86-755-86028224
Fax. + 86-755-26654850



Lotus Global Co., Ltd
15 Alexandra Road
London
NW8 0DP
UK
Tel. + 44-20-75868010
Fax. + 44-20-79006187



FOR PROFESSIONAL USE ONLY
Store at 2...8 °C



**COMPLETELY READ THE INSTRUCTIONS BEFORE
PROCEEDING**

en

SYMBOLS EXPLANATIONS



Authorized Representative in Europe



Manufacturer



Attention. See Instructions For Use



Contents of kit



In vitro diagnostic medical device
(In vitro diagnostic use)



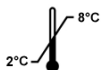
Lot number



Catalogue Code



Expiry date (Use by...)



Temperature limitation
(store at 2...8 °C)



Number of tests



Keep away from sunlight



Keep upright

INTENDED USE

The kit has been designed for the qualitative determination of Rubella IgG in human serum

The method can be used for samples over the range of 0-30 AU/ml.

The test has to be performed on the MAGLUMI chemiluminescence immunoassay (CLIA) fully auto analyzer (Including MAGLUMI 1000, MAGLUMI 2000, MAGLUMI 2000 Plus and new developed models).

SUMMARY AND EXPLANATION OF THE TEST

Rubella is a viral exanthematous infectious disease caused by rubella virus, a single-stranded RNA virus belonging to the Togavirus group. The illness follows a typically benign clinical course with rare complications and is subclinical in a large proportion of cases. Symptomatology is generally mild, characterized by fever, malaise, a maculopapular rash of three to five days' duration and, possibly, coryza and conjunctivitis. The disease is usually accompanied by lymphadenopathy. Infection confers lifelong immunity.

Infection from rubella virus is particularly disastrous if contracted during the first four months of gestation. If not immunologically protected, women infected during pregnancy run a high risk of embryofetal damage. Congenital rubella causes a wide range of severe defects, many of which are permanent and adversely affect later development (cataract, deafness, hepatosplenomegaly, psychomotor retardation, bone alterations, cardiopathies, and neuropathies). Pathological consequences on the foetus or newborn depend on teratogenicity of the virus and on the time of pregnancy when the infection has been contracted. Gestational age at the time of maternal infection is considered the most important determinant of intrauterine transmission and foetal damage. It is generally accepted that the risk decreases with increasing gestational age: it is highest in case of infection during the first two months of pregnancy (40-60%) and progressively decreases during the fourth and fifth months (10-20%). Clinical findings in newborns and virus isolation studies have demonstrated that foetal infection is rare beyond the second trimester of gestation.

Rubella virus is transmitted in utero during the course of primary maternal infection, whether apparent or inapparent, when the virus in the bloodstream infects the placenta and, subsequently, the foetus. Intrauterine transmission of virus associated with maternal re-infection is extremely rare, indicating that maternal immunity (whether naturally derived or vaccine-induced) protects against intrauterine infection. Maternal infection may result in (a) no infection of the embryo; (b) resorption of the embryo (seen only with infections occurring in the earliest stages of gestation); (c) miscarriage; (d) stillbirth; (e) infection of placenta without foetal involvement or (f) infection of both the placenta and foetus. Infected infants may present obvious multiple organ involvement or, as is frequently observed, no immediately evident disease. However, after long-term follow-up, many of these seemingly unaffected infants have evidence of hearing loss, or central nervous system lesions, or other defects.

The first humoral immune response to infection is the synthesis of specific anti-rubella virus IgM antibody which reaches high serum levels two weeks after the rash and lasts in the circulation for one to two month(s). Specific IgG antibody generally appears a few days after the onset of rash, about one week after IgM develops. It rapidly increases to reach a plateau six to ten weeks after the onset of symptoms and then progressively decreases to a level (15-200 IU/mL) lasting for the whole life. Reinfection, completely asymptomatic, is accompanied by moderately increased levels of

specific IgG.

Correct detection of IgM and IgG antibodies to rubella virus provides an essential tool for diagnosing and following up acute infection, for assessment of immune status in fertile women, and therefore for adopting suitable prophylaxis in susceptible women of child-bearing age. Since when a vaccine was made available, the assay of IgG to rubella virus has been widely used to determine seroconversion of the recipient after vaccination.

PRINCIPLE OF THE TEST

Indirect immunoluminometric assay;

Mouse anti-human IgG is used to label ABEI, and use purified Rubella antigen to coat nano magnetic microbeads. Sample, Calibrator or Control with Buffer and nano magnetic microbeads coated with Rubella antigen are mixed thoroughly and incubated at 37 °C and cycle washing for 1 time. Then add ABEI Label, incubation and form a sandwich, then washing for the 2nd time. Subsequently, the starter reagents are added and a flash chemiluminescent reaction is initiated. The light signal is measured by a photomultiplier as RLU within 3 seconds and is proportional to the concentration of Rubella IgG present in controls or samples.

CONT

KIT COMPONENTS

Material Supplies

Reagent Integral for 100 determinations	
Nano magnetic microbeads: TRIS buffer, 1.2% (W/V), 0.2%NaN ₃ , coated with Rubella antigen	2.5ml
Calibrator Low: bovine serum, 0.2%NaN ₃ .	2.5ml
Calibrator High: vine serum, 0.2%NaN ₃	2.5ml
Buffer: sheep anti-human IgA, sheep anti-human IgM containing BSA, 0.2%NaN ₃ .	12.5ml
ABEI Label: Mouse anti-human IgG monoclonal antibody labeled ABEI, contains BSA, 0.2%NaN ₃	22.5ml
Diluent: sheep anti-human IgA, sheep anti-human IgM containing BSA, 0.2%NaN ₃ .	25ml
All reagents are provided ready-to-use.	

Please prepare 0.9% sodium chloride solution in case of insufficient diluents.

Reagent Vials in kit box	
Internal Quality Control: containing BSA, 0.2%NaN ₃ . (target value refer to Quality Control Information date sheet)	2.0ml

Accessories Required But Not Provided

MAGLUMI Reaction Module	REF: 630003
MAGLUMI Starter 1+2	REF: 130299004M
MAGLUMI Wash Concentrate	REF: 130299005M
MAGLUMI Light Check	REF: 130299006M



Preparation of the Reagent Integral

Before the sealing is removed, gentle and careful horizontal shaking of the Reagent Integral is essential (avoid foam formation!) Remove the sealing and turn the small wheel of the magnetic microbeads compartment to and fro, until the colour of the suspension has changed into brown. Place the Integral into the reagent area and let it stand there for 30 min. During this time, the magnetic microbeads are automatically agitated and completely resuspended.

Do not interchange integral component from different reagents or lots!

076120522-v1.0-EN

Storage and Stability

- Sealed: Stored at 2-8°C until the expiry date.
- Opened: Stable for 4 weeks. To ensure the best kit performance, it is recommended to place opened kits in the refrigerator if it's not going to be used on board during the next 12 hours.



- Keep upright for storage.



- Keep away from direct sunlight.

CALIBRATION AND TRACEABILITY

1) Traceability

To perform an accurate calibration, we have provided the test calibrators standardized against the SNIBE internal reference substance.

Calibrators in the Reagent Kit are from Fitzgerald.

2) 2-Point Recalibration

Via the measurement of calibrators, the predefined master curve is adjusted (recalibrated) to a new, instrument-specific measurement level with each calibration.

3) Frequency of Recalibration

- After each exchange of lots (Reagent Integral or Starter Reagents).
- Every 4 weeks and/or each time a new Integral is used (recommendation).
- After each servicing of the MAGLUMI Fully Auto analyzer.
- If controls are beyond the expected range.

SPECIMEN COLLECTION AND PREPARATION

Sample material: serum

Collect samples using standard procedures.

Store at 2-8°C: 24 hours, for longer storage periods: freeze to below -20°C

Avoid repeated freezing and thawing cycles, stored samples should be thoroughly mixed prior to use (Vortex mixer).

Please ask local representative of SNIBE for more details if you have any doubt.

Vacuum Tubes

- (a) Blank tubes are recommended type for collecting samples.
- (b) Please ask SNIBE for advice if special additive must be used in sample collecting.

Specimen Conditions

- Do not use specimens with the following conditions:
 - (a) heat-inactivated specimens;
 - (b) Cadaver specimens or body fluids other than human serum;
 - (c) Obvious microbial contamination.
- Use caution when handling patient specimens to prevent cross contamination. Use of disposable pipettes or pipette tips is recommended.
- Inspect all samples for bubbles. Remove bubbles with an applicator stick prior to analysis. Use a new applicator stick for each sample to prevent cross contamination.
- Serum specimens should be free of fibrin, red blood cells or other particulate matter.
- Ensure that complete clot formation in serum specimens has taken place prior to centrifugation. Some specimens, especially those from patients receiving anticoagulant or thrombolytic therapy, may exhibit increased clotting time. If the specimen is centrifuged before a complete clot forms, the presence of fibrin may cause erroneous results.

Preparation for Analysis

- Patient specimens with a cloudy or turbid appearance must be centrifuged prior to testing. Following centrifugation, avoid the lipid layer (if present) when pipetting the specimen into a sample cup or secondary tube.
- Specimens must be mixed **thoroughly** after thawing by **low** speed vortexing or by gently inverting, and centrifuged prior to use to remove red blood cells or particulate matter to ensure consistency in the results. Multiple freeze-thaw cycles of specimens should be avoided.
- All samples (patient specimens or controls) should be tested within 3 hours of being placed on board the MAGLUMI System. Refer to the SNIBE service for a more detailed discussion of onboard sample storage constraints.

Storage

- If testing will be delayed for more than 8 hours, remove serum from the serum separator, red blood cells or clot. Specimens removed from the separator gel, cells or clot may be stored up to 24 hours at 2-8°C.
- Specimens can be stored up to 30 days frozen at -20°C or colder.

Shipping

- Before shipping specimens, it is recommended that specimens be removed from the serum or plasma separator, red blood cells or clot. When shipped, specimens must be packaged and labeled in compliance with applicable state, federal and international regulations covering the transport of clinical specimens and infectious substances. Specimens must be shipped frozen (dry ice). Do not exceed the storage time limitations identified in this section of the package insert.

WARNING AND PRECAUTIONS FOR USERS



- For use in *IN-VITRO* diagnostic procedures only.
- Package insert instructions must be carefully followed. Reliability of assay results cannot be guaranteed if there are any deviations from the instructions in this package insert.

Safety Precautions

CAUTION: This product requires the handling of human specimens.

- The calibrators in this kit are prepared from bovine serum products. However, because no test method can offer complete assurance that HIV, Hepatitis B Virus or other infectious agents are absent; these reagents should be considered a potential biohazard and handled with the same precautions as applied to any serum or plasma specimen.
- All samples, biological reagents and materials used in the assay must be considered potentially able to transmit infectious agents. They should therefore be disposed of in accordance with the prevailing regulations and guidelines of the agencies holding jurisdiction over the laboratory, and the regulations of each country. Disposable materials must be incinerated; liquid waste must be decontaminated with sodium hypochlorite at a final concentration of 5% for at least half an hour. Any materials to be reused must be autoclaved using an overkill approach (USP 24, 2000, p.2143). A minimum of one hour at 121 °C is usually considered adequate, though the users must check the effectiveness of their decontamination cycle by initially validating it and routinely using biological indicators.
- It is recommended that all human sourced materials be considered potentially infectious and handled in accordance with the OSHA Standard on Bloodborne Pathogens¹³. Biosafety Level 214 or other appropriate biosafety practices should be used for materials that contain or are suspected

of containing infectious agents.

- This product contains Sodium Azide; this material and its container must be disposed of in a safe way.
- Safety data sheets are available on request.

Handling Precautions

- Do not use reagent kits beyond the expiration date.
- Do not mix reagents from different reagent kits.
- Prior to loading the Reagent Kit on the system for the first time, the microbeads requires mixing to re-suspend microbeads that have settled during shipment.
- For microbeads mixing instructions, refer to the KIT COMPONENTS, Preparation of the Reagent Integral section of this package insert.
- To avoid contamination, wear clean gloves when operating with a reagent kit and sample.
- Over time, residual liquids may dry on the kit surface, please pay attention the silicon film still exists on the surface of the kit.
- For a detailed discussion of handling precautions during system operation, refer to the SNIBE service information.

TEST PROCEDURE

To ensure proper test performance, strictly adhere to the operating instructions of the MAGLUMI Fully Auto analyzer. Each test parameter is identified via a RFID tag on the Reagent Integral. For further information please refer to the MAGLUMI Chemiluminescence Analyzer Operating Instructions.

Auto-dilution 1:11 20µl +200µl	Sample Diluent
20µl +100µl +20µl	Diluted Sample, calibrator or controls Buffer Nano magnetic microbeads
10 min	Incubation
400µl	Cycle washing
+200µl	ABEI Label
10 min	Incubation
400µl	Cycle washing
3 s	Measurement

* In case of lacking diluent, user can prepare 0.9% sodium chloride solution as additional diluent.

DILUTION

Samples with concentrations above the measuring range can be diluted. After manual dilution, multiply the result by the dilution factor. After dilution by the analyzers, the analyzer software automatically takes the dilution into account when calculating the sample concentration.

Availability of sample dilution by analyzer please refers to the MAGLUMI analyzer user software program. Dilution settings please follow MALGUMI analyzer operating instructions.

QUALITY CONTROL

- Observe quality control guidelines for medical laboratories
- Use suitable controls for in-house quality control. Controls should be run at least once every 24 hours when the test is in use, once per reagent kit and after every calibration. The control intervals should be adapted to each laboratory's individual requirements. Values obtained should fall within the defined ranges. Each laboratory should establish guidelines for corrective measures to be taken if values fall outside the range.

LIMITATIONS OF THE PROCEDURE

1) Limitations

Use Rubella IgG value as a kind of auxiliary material for other testing data when in diagnosis. Assay results should be utilized in conjunction with other clinical and laboratory data to assist the

clinician in making individual patient management decisions
A skillful technique and strict adherence to the instructions are necessary to obtain reliable results. Bacterial contamination of samples or repeated freeze-thaw cycles may affect the test results. Assay results should be utilized in conjunction with other clinical and laboratory data to assist the clinician in making individual patient management decisions.

2) Interfering Substances

No interference with test results is seen by concentrations of bilirubin < 0.06mg/ml, haemoglobin < 16mg/dl or triglycerides < 12.5mg/ml.

3) HAMA

Patient samples containing human anti-mouse antibodies (HAMA) may give falsely elevated or decreased values. Although HAMA-neutralizing agents are added, extremely high HAMA serum concentrations may occasionally influence results.

RESULTS

1) Calculation of Results

- The analyzer automatically calculates the Rubella IgG concentration in each sample by means of a calibration curve which is generated by a 2-point calibration master curve procedure. The results are expressed in AU/ml. For further information please refer to the MAGLUMI Chemiluminescence Analyzer Operating Instructions.
- Test results need NOT to multiply dilution rate!**

2) Interpretation of Results

Results obtained with the MAGLUMI Rubella IgG assay can be interpreted as follows:

- Non-reactive: A result less than 2 AU/ml (< 2 AU/ml) is considered to be negative.
- Reactive: A result greater than or equal to 2 AU/ml (≥ 2 AU/ml) considered to be positive.

Results from assays of other manufacturers cannot be used interchangeably.

PERFORMANCE CHARACTERISTICS

1) Precision

Intra-assay coefficient of variation was evaluated on 3 different levels of control serum repeatedly measured 20 times in the same run, calculating the coefficient of variation.

Intra-assay precision			
Control	Mean(AU/ml)	SD(AU/ml)	CV%
Level 1	1.61	0.09	5.86
Level 2	8.15	0.48	5.89
Level 3	19.55	1.10	5.65

Inter-assay coefficient of variation was evaluated on three batches of kits. Repeatedly measured 3 different levels of control serum 21 times, calculating the coefficient of variation.

Inter-assay precision			
Control	Mean(AU/ml)	SD(AU/ml)	CV%
Level 1	1.57	0.14	8.85
Level 2	8.24	0.70	8.55
Level 3	19.24	1.59	8.26

2) Analytical Sensitivity

The sensitivity is defined as the concentration of Rubella IgG equivalent to the mean RLU of 20 replicates of the zero standard plus two standard deviations corresponding to the concentration from the standard curve. The sensitivity is typically less than 0.25 AU/ml.

3) Specificity

The specificity of the Rubella IgG assay system was assessed by measuring the apparent response of the assay to various potentially cross reactive analytes:

When CMV IgG, CMV IgM, Rubella IgM, Toxo IgG, Toxo IgM, HSV-1/2IgG, HSV-1/2IgM separately reach a concentration of 30AU/ml, measured Toxo IgG is negative. No cross reaction with the IgG or IgM antibody of HAV, HBV, HCV, HIV, syphilis, EBV. The ELISA diagnosed RF or ANA positive, which is non Rubella infected sample, this reagent's determination results show negative.

4) Recovery

Consider calibrator high of known concentration as a sample, dilute it by 1:2 ratio with diluents, and measure its diluted concentration for 10 times. Then calculate the recovery of measured concentration and expected concentration. The recovery should be within 90% -110%.

Expected	Mean Measuring	Recovery
9.8 AU/ml	9.9 AU/ml	101%

5) Linearity

Use Rubella IgG calibrator to prepare the six-point standard curve, measuring all points' RLU except point A, and then do four-parameter linear fitting in double logarithm coordinate, the absolute linear correlation coefficient(r) should be bigger than 0.9800.

Calibrator Point	Concentration AU/ml	Absolute linear correlation coefficient (r)
A	0.0	
B	1.0	r=0.9847
C	3.0	
D	8.0	
E	15.0	
F	30.0	

REFERENCES

- Lisa Byrne, Lisa Brant, Claire Reynolds, Mary Ramsay, Vaccine, Volume 30, Issue 2, 5 January 2012, Pages 161-167.
- Blenda Böttiger, Inge Panum Jensen, Clinical and Diagnostic Virology, Volume 8, Issue 2, August 1997, Pages 105-111.
- Mohammed-Durosinlorun Amina, Shittu Oladapo, Sadauki Habib, Olayinka Adebola, Kolawole Bimbo, Adejo Daniel, International Health, Volume 2, Issue 2, June 2010, Pages 156-159.
- H.I.J. Thomas, P. Morgan-Capner, Journal of Virological Methods, Volume 31, Issues 2-3, February-March 1991, Pages 219-228.
- Klaus-Peter Wandinger, Sandra Saschenbrecker, Katja Steinhagen, Thomas Scheper, Wolfgang Meyer, Uwe Bartelt, Gisela Enders, Journal of Virological Methods, Volume 174, Issues 1-2, June 2011, Pages 85-93.
- Giuseppe Portella, Claudio Galli, for the MIGHT (Multicenter Italian Group for Hospital ToRCH evaluation), Journal of Clinical Virology, Volume 49, Issue 2, October 2010, Pages 105-110.
- A. Paris-Hamelin, S. Fustec-Ibarboure, Journal of Virological Methods, Volume 10, Issue 4, April 1985, Pages 355-361
- Best, J. M., S. O'Shea, G. Tipples, N. Davies, S. M. Al Khusaiby, A. Krause, L. M. Hesketh, L. Jin, and G. Enders. 2002. Interpretation of rubella serology in pregnancy—pitfalls and problems. *BMJ* 325:147-148.
- Dwyer, D. E., P. W. Robertson, and P. R. Fields. 2001. Broadsheet: clinical and laboratory features of rubella. *Pathology* 33:322-328.
- Field, P. R., V. Sintchenko, and G. L. Gilbert. 1998. Confirmation of rubella infection: review of 10 years' experience at ICPMR. *Inoculum* 7:1-3.
- Francis, B. H., A. K. Thomas, and C. A. McCarty. 2003. The impact of rubella immunization on the serological status of

women of childbearing age: a retrospective longitudinal study in Melbourne, Australia. *Am. J. Public Health* 93:1274-1276

12. Gutiérrez, A., M. J. Rodríguez, F. De Ory, G. Piédrola, and M. C. Maroto. 1999. Reliability of low-avidity IgG and of IgA in the diagnosis of primary infection by rubella virus with an adaptation of a commercial test. *J. Clin. Lab. Anal.* 13:1-4.