MAGLUMI CEA (CLIA)



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Shenzhen New Industries Biomedical Engineering Co., Ltd 4F,Wearnes Tech Bldg, Science & Industry Park, Nanshan,Shenzhen,518057CHINA Tel. + 86-755-86028224 Fax.+ 86-755-26654850 Lotus Global Co., Ltd 15 Alexandra Road London NW8 0DP UK Tel. + 44-20-75868010 Fax.+ 44-20-79006187

CE

FOR PROFESSIONAL USE ONLY

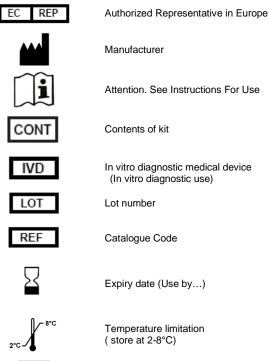
Store at 2-8°C



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COMPLETELY READ THE INSTRUCTIONS BEFORE PROCEEDING

SYMBOLS EXPLANATIONS





Keep away from direct sunlight

Number of tests

Keep upright for storage

INTENDED USE

The kit has been designed for the quantitative determination of Carcinoembryonic Antigen (CEA) in human serum.

The method can be used for samples over the range of 0-1,000 ng/ml.

The test has to be performed on the MAGLUMI chemiluminescence immunoassay (CLIA) fully auto analyzer (Including MAGLUMI 1000, MAGLUMI 2000, MAGLUMI 2000 Plus and new developed models).

SUMMARY AND EXPLANATION OF THE TEST

CEA was first isolated from colon tumors. It belongs to a family of glycoproteins with a molecular weight of approx. 180,000-200,000 and a carbohydrate content varying between 50 and 60%.

Elevated CEA serum concentrations are found in patients with carcinoma of the lung, colon or breast.

CEA measurement, alone or in combination with other markers, is intended for early diagnosis of relapses for therapy monitoring.

PRINCIPLE OF THE TEST

Sandwich immunoluminometric assay:

Use an anti-CEA monoclonal antibody to label ABEI, and use another monoclonal antibody to label FITC. Sample, Calibrator or Control are mixed thoroughly with FITC Label and nano magnetic microbeads in a cuvette incubated at 37°C, then cycle washing for 1 time. Then add ABEI Label and incubated to form a sandwich, after sediment in a magnetic field, suck the supernatant then cycle washing for the 2nd time. Subsequently, Starter1+2 substrates are added and a flash chemiluminescent reaction is initiated. The light signal is measured by a photomultiplier as RLU within 3 seconds and is proportional to the concentration of CEA present in controls or samples.



KIT COMPONENTS

Material supplies

Reagent Integral for 100 determinations	
Nano magnetic microbeads: microbeads	
coated with sheep anti-FITC polyclonal	2.5ml
antibody, TRIS buffer, 0.2%NaN ₃ .	
Calibrator Low: bovine serum, 0.2%NaN ₃ .	2.5ml
Calibrator High: bovine serum, 0.2%NaN ₃	2.5ml
FITC Label: anti-CEA monoclonal antibody 12.5ml	
labeled FITC, containing BSA, 0.2%NaN ₃ .	12.5111
ABEI Label: anti-CEA monoclonal antibody 22.5ml	
labeled ABEI, containing BSA, 0.2%NaN ₃ .	22.500
Diluent: 0.9%NaCl. 25ml	
All reagents are provided ready-to-use.	

Reagent Vials in kit box		
Internal Quality Control: containing BSA,		
0.2%NaN ₃ . (target value refer to Quality	2.0ml	
Control Information date sheet)		

Accessories Required But Not Provided

MAGLUMI Reaction Module	REF: 630003
MAGLUMI Starter1+2	REF: 130299004M
MAGLUMI Wash Concentrate	REF: 130299005M
MAGLUMI Light Check	REF: 130299006M



Before the sealing is removed, gently and carefully horizontally shaking of the Reagent Integral is essential (avoid foam formation!). Remove the sealing and turn the small wheel of the magnetic microbeads compartment to and fro, until the color of the suspension has changed into brown. Place the Integral into the reagent area and let it stand there for 30min. During this time, the magnetic microbeads are automatically agitated and completely resuspended.

Do not interchange Nano Magnetic Microbeads from different reagents or lots!

Storage and Stability

- Sealed: Stored at 2-8°C until the expiry date.
- Opened: Stable for 4 weeks. To ensure the best kit performance, it is recommended to place opened kits in the refrigerator if it's not going to be used on board during the next 12 hours.

Keep upright for storage

TRACEABILITY AND CALIBRATION

1)Traceability

To perform an accurate calibration, we have provided the test calibrators standardized against the W.H.O.1st International Reference Preparation 73/601.

2) 2-Point Recalibration

Via the measurement of calibrators, the predefined master curve is adjusted (recalibrated) to a new, instrument-specific measurement level with each calibration.

3) Frequency of Recalibration

- After each exchange of lots (Reagent Integral or Starter Reagents).
- Every 4 weeks and/or each time a new Integral is used (recommendation).
- After each servicing of the MAGLUMI Fully Auto analyzer.
- If controls are beyond the expected range.

SPECIMEN COLLECTION AND PREPARATION

Sample material: serum

Collect samples using standard procedures.

Store at 2-8°C: 24 hours, for longer storage periods: freeze to below - 20° C.

Avoid repeated freezing and thawing cycles, stored samples should be thoroughly mixed prior to use (Vortex mixer).

Please ask local representative of SNIBE for more details if you have any doubt.

Vacuum Tubes

(a) Blank tubes are recommended type for collecting samples.(b) Please ask SNIBE for advice if special additive must be used in sample collecting.

Specimen Conditions

- · Do not use specimens with the following conditions:
- (a) heat-inactivated specimens;
- (b) Cadaver specimens or body fluids other than human serum;
- (c) Obvious microbial contamination.
- Use caution when handling patient specimens to prevent cross contamination. Use of disposable pipettes or pipette tips

is recommended.

- Inspect all samples for bubbles. Remove bubbles with an applicator stick prior to analysis. Use a new applicator stick for each sample to prevent cross contamination.
- Serum specimens should be free of fibrin, red blood cells or other particulate matter.
- Ensure that complete clot formation in serum specimens has taken place prior to centrifugation. Some specimens, especially those from patients receiving anticoagulant or thrombolytic therapy, may exhibit increased clotting time. If the specimen is centrifuged before a complete clot forms, the presence of fibrin may cause erroneous results.

Preparation for Analysis

- Patient specimens with a cloudy or turbid appearance must be centrifuged prior to testing. Following centrifugation, avoid the lipid layer (if present) when pipetting the specimen into a sample cup or secondary tube.
- Specimens must be mixed thoroughly after thawing by low speed vortexing or by gently inverting, and centrifuged prior to use to remove red blood cells or particulate matter to ensure consistency in the results. Multiple freeze-thaw cycles of specimens should be avoided.
- All samples (patient specimens or controls) should be tested within 3 hours of being placed on board the MAGLUMI System. Refer to the SNIBE service for a more detailed discussion of onboard sample storage constraints.

Storage

- If testing will be delayed for more than 8 hours, remove serum or plasma from the serum or plasma separator, red blood cells or clot. Specimens removed from the separator gel, cells or clot may be stored up to 24 hours at 2-8°C.
- Specimens can be stored up to 30 days frozen at -20°C or colder.

Shipping

Before shipping specimens, it is recommended that specimens be removed from the serum or plasma separator, red blood cells or clot. When shipped, specimens must be packaged and labeled in compliance with applicable state, federal and international regulations covering the transport of clinical specimens and infectious substances. Specimens must be shipped frozen (dry ice). Do not exceed the storage time limitations identified in this section of the package insert.

WARNING AND PRECAUTIONS FOR USERS



- For use in *IN-VITRO* diagnostic procedures only.
- Package insert instructions must be carefully followed. Reliability of assay results cannot be guaranteed if there are any deviations from the instructions in this package insert.

Safety Precautions

CAUTION: This product requires the handling of human specimens.

- The calibrators in this kit are prepared from bovine serum products. However, because no test method can offer complete assurance that HIV, Hepatitis B Virus or other infectious agents are absent; these reagents should be considered a potential biohazard and handled with the same precautions as applied to any serum or plasma specimen.
- All samples, biological reagents and materials used in the assay must be considered potentially able to transmit infectious agents. They should therefore be disposed of in accordance with the prevailing regulations and guidelines of the agencies holding jurisdiction over the laboratory, and the

regulations of each country. Disposable materials must be incinerated; liquid waste must be decontaminated with sodium hypochlorite at a final concentration of 5% for at least half an hour. Any materials to be reused must be autoclaved using an overkill approach (USP 24, 2000, p.2143). A minimum of one hour at 121°C is usually considered adequate, though the users must check the effectiveness of their decontamination cycle by initially validating it and routinely using biological indicators.

- It is recommended that all human sourced materials be considered potentially infectious and handled in accordance with the OSHA Standard on Blood borne Pathogens13. Biosafety Level 214 or other appropriate biosafety practices should be used for materials that contain or are suspected of containing infectious agents.
- This product contains Sodium Azide; this material and its container must be disposed of in a safe way.
- Safety data sheets are available on request.

Handling Precautions

- Do not use reagent kits beyond the expiration date.
- · Do not mix reagents from different reagent kits.
- Prior to loading the Reagent Kit on the system for the first time, the microbeads requires mixing to re-suspend microbeads that have settled during shipment.
- For microbeads mixing instructions, refer to the KIT COMPONENTS, Preparation of the Reagent Integral section of this package insert.
- To avoid contamination, wear clean gloves when operating with a reagent kit and sample.
- Over time, residual liquids may dry on the kit surface; please pay attention to the silicon film still exists on the surface of the kit.
- For a detailed discussion of handling precautions during system operation, refer to the SNIBE service information.

TEST PROCEDURE

To ensure proper test performance, strictly adhere to the operating instructions of the MAGLUMI Fully Auto analyzer. Each test parameter is identified via a RFID tag on the Reagent Integral. For further information please refer to the MAGLUMI Chemiluminescence Analyzer Operating Instructions.

40µl	Sample, calibrator or controls	
+100µl	FITC label	
+20µl	Nano magnetic microbeads	
10min	Incubation	
400µl	Cycle washing	
+200µl	ABEI label	
10 min	Incubation	
400µl	Cycle washing	
3 s	Measurement	

DILUTION

Samples with concentrations above the measuring range can be diluted. After manual dilution, multiply the result by the dilution factor. After dilution by the analyzers, the analyzer software automatically takes the dilution into account when calculating the sample concentration.

Availability of sample dilution by analyzer please refers to the MAGLUMI analyzer user software program. Dilution settings please follow MAGLUMI analyzer operating instructions.

QUALITY CONTROL

- · Observe quality control guidelines for medical laboratories.
- Use suitable controls for in-house quality control. Controls should be run at least once every 24 hours when the test is in use, once per reagent kit and after every calibration. The control intervals should be adapted to each laboratory's

individual requirements. Values obtained should fall within the defined ranges. Each laboratory should establish guidelines for corrective measures to be taken if values fall outside the range.

LIMITATIONS OF THE PROCEDURE

1) Limitations

Patients with malignancies may exhibit CEA values within the reference range. Elevated CEA values may also be observed in patients with benign diseases such as liver cirrhosis, viral hepatitis, pancreatic or gastrointestinal disorders. Smoking and alcohol consumption may also lead to an increase in CEA concentrations. Therefore, CEA serum levels may only be interpreted in context with the clinical picture and other diagnostic procedures.

2) Interfering Substances

No interference with test results is seen by concentrations of bilirubin<0.125mg/ml, haemoglobin<16mg/dl or triglycerides<12.5 mg/ml.

3) HAMA

Patient samples containing human anti-mouse antibodies (HAMA) may give falsely elevated or decreased values. Although HAMA-neutralizing agents are added, extremely high HAMA serum concentrations may occasionally influence results.

4) High-Dose Hook

High dose hook is a phenomenon whereby very high level specimens may read within the dynamic range of the assay. For the MAGLUMI CEA assay, no high dose hook effect was observed when samples containing up to 200,000ng/ml.

RESULTS

1) Calculation of Results

- The analyzer automatically calculates the CEA concentration in each sample by means of a calibration curve which is generated by a 2-point calibration master curve procedure. The results are expressed in ng/ml. For further information please refer to the MAGLUMI Chemiluminescence Analyzer Operating Instructions.
- Conversion factor: 1ng/ml = 15mIU/ml.

2) Interpretation of Results

- Results of study in clinical centers with group of individuals, 95% of the results were< 5.093ng/ml.
- Results may differ between laboratories due to variations in population and test method. If necessary, each laboratory should establish its own reference range.

PERFORMANCE CHARACTERISTICS

1) Precision

Intra-assay coefficient of variation was evaluated on 3 different levels of control serum repeatedly measured 20 times in the same run, calculating the coefficient of variation.

Intra-assay precision			
Control	Mean(ng/ml)	SD(ng/ml)	CV%
Level 1	15.31	0.97	6.22
Level 2	22.54	1.32	5.87
Level 3	45.92	2.45	5.34

Inter-assay coefficient of variation was evaluated on three batches of kits. Repeatedly measured 3 different levels of control serum 21 times, calculating the coefficient of variation.

Inter-assay precision CV% Control Mean(ng/ml) SD(ng/ml) Level 1 15.48 1.74 9.49 Level 2 23.73 2.32 9.76 4.03 Level 3 44.15 9.13

2) Analytical Sensitivity

The sensitivity is defined as the concentration of CEA equivalent to the mean RLU of 20 replicates of the zero standard plus two standard deviations corresponding to the concentration from the standard curve. The sensitivity is typically less than 0.5ng /ml.

3) Specificity

The specificity of the CEA assay system was assessed by measuring the apparent response of the assay to various potentially cross reactive analytes.

Compound	Concentration	Cross reactivity
AFP	100 IU/ml	1%
CA125	100 IU/mI	0.8%
CA153	100 IU/ml	0.9%

4) Recovery

Consider calibrator high of known concentration as a sample, dilute it by 1:2 ratios with diluents, and measure its diluted concentration for 10 times. Then calculate the recovery of measured concentration and expected concentration. The recovery should be within 90% -110%.

Expected	Mean Measuring	Recovery
210.85ng/ml	213.85ng/ml	103%

5) Linearity

Use CEA calibrator to prepare the six-point standard curve, measuring all points' RLU except point A, and then do four-parameter linear fitting in double logarithm coordinate, the absolute linear correlation coefficient(r) should be bigger than 0.9800.

Calibrator Point	Concentration ng/ml	Absolute linear correlation coefficient (r)
A	0	
В	5.333	
С	13.333	0.9910
D	33.333	
E	66.666	
F	166.666	

6) Method comparison

A comparison of MAGLUMI CEA (y) with a commercially available CEA test (x) using clinical samples gave the following correlations (ng/ml):

Linear regression y = 1.08x-10.34

r = 0.967 Sy.x = 18.32

Number of samples measured: 173

The sample concentrations were between 3.74 and 2000.45 $\ensuremath{\text{ng/ml}}$

REFERENCES

- Chu DZJ, Erickson CA, Russel, MP, Thompson C, Lang NP, Broadwater RJ, Westbrook KC. Prognostic Significance of Carcinoembryonic Antigen in Colorectal Carcinoma. Arch Surg 1991; 126: 314-316.
- Díez M, Torres A, Maestro ML, Ortega MD, Gómez A, Pollán M, Lopez JA, Picardo A, Hernando F, Balibrea JL. Prediction of survival and recurrence by serum and cytosolic levels of CEA, CA125 and SCC antigens in resectable non-small-cell lung cancer. Br J Cancer 1996; 73; 1248-1254.
- Dnistrian AM Schwartz MK, Greenberg EJ, Smith CA, Schwartz DC. CA 15-3 and carcinoembryonic antigen in the clinical evaluation of breast cancer. Clinical Chimica Acta 1991; 200: 81-94.

4. Gold P, Freedman SO. Demonstration of Tumor-Specific 025120530-v1.0-EN

Antigens in Human Colonic Carcinomata by Immunological Tolerance and Absorption Techniques. J Exp Med 1965; 121: 439-462.

- 5. Kuroki M, Arakawa F, Higuchi H, Matsunaga A, Okamoto N, Takakura K, Matsuoka Y. Epitope Mapping of the Carcinoembryonic Antigen by Monoclonal Antibodies and Establishment of a New Improved Radioimmunoassay System. Jpn J Cancer Res (Gann) 1987; 78: 386-396.
- 6. Lamerz R. CEA in Clinical Practice. J Nucl Med Allied Sci 1990; 34 (Suppl. 3): 41-48.
- Murakami M, Kuroki M, Arakawa F, Kuwahara M, Oikawa S, Nakazato, H, Matsuoka Y. A Reference of the GOLD Classification of Monoclonal Antibodies against Carcinoembryonic Antigen to the Domain Structure of the Carcinoembryonic Antigen Molecule. Hybridoma 1995; 14 (1): 19- 28.
- Sikorska HM, Fuks A, Gold P. Clinical Applications of Carcinoembryogenic Antigen. J Nucl Med Allied Sci 1990; 34 (Suppl. 3): 7-29.
- 9. Von Kleist S, Zimmermann W, Thompson J. The Carcinoembryonic Gene Family. J Nucl Med Allied Sci 1990; 34 (Suppl. 3): 30-33.